



The Role of Glial Mitochondria in α-Synuclein Toxicity

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The abnormal accumulation of alpha-synuclein (α -syn) aggregates in neurons and glial cells is widely known to be associated with many neurodegenerative diseases, including Parkinson's disease (PD), Dementia with Lewy bodies (DLB), and Multiple system atrophy (MSA). Mitochondrial dysfunction in neurons and glia is known as a key feature of α -syn toxicity. Studies aimed at understanding α -syn-induced toxicity and its role in neurodegenerative diseases have primarily focused on neurons. However, a growing body of evidence demonstrates that glial cells such as microglia and astrocytes have been implicated in the initial pathogenesis and the progression of α -Synucleinopathy. Glial cells are important for supporting neuronal survival, synaptic functions, and local immunity. Furthermore, recent studies highlight the role of mitochondrial metabolism in the normal function of glial cells. In this work, we review the complex relationship between glial mitochondria and α -syn-mediated neurodegeneration, which may provide novel insights into the roles of glial cells in α -syn-associated neurodegenerative diseases.

Keywords: alpha-synuclein, mitochondrial dysfuncion, neurodegenerative diseases, non-cell autonomous, glia, neuron

INTRODUCTION

Alpha synuclein (α -syn), first identified in 1988 by Maroteaux et al. (1988), is a small protein that consists of 140 amino acids and encodes the human SNCA gene. α -syn is expressed in the central nervous system (CNS), and is specifically localized in synapses and nuclei (Jakes et al., 1994). The function of α -syn is not clearly defined, but several studies have shown that α -syn regulates synaptic plasticity and vesicle trafficking (Kahle et al., 2000; Fortin et al., 2004; Lee et al., 2008) and interacts with synaptic vesicles to physiologically regulate vesicle recycling (Maroteaux et al., 1988; Huang et al., 2019). Originally, α -syn was thought to be a natively unfolded monomeric protein, but recent studies have indicated that α -syn forms an α -helix-rich tetramer (Bartels et al., 2011; Wang et al., 2011, 2014; Dettmer et al., 2013, 2015; Gurry et al., 2013; Burre et al., 2014). The biological function of α -syn is exerted through the N-terminal, non-amyloid-beta component (NAC) and C-terminal domains. The N-terminus, which contains the KTKEGV motif, maintains tetramerization of α -syn, and mutations in this motif can induce neurotoxicity (Dettmer et al., 2015). NAC, first identified in Alzheimer's disease patients, is a highly hydrophobic domain and forms a β -sheet structure for α -syn aggregation (Ueda et al., 1993). The C-terminus of α -syn is a proline-rich region. α -syn can interact with other proteins through this domain (Kim et al., 2002). Unfolded or misfolded α -syn forms fibrillar aggregates and aggregated α -syn generates insoluble inclusions in the affected neurons and glial cells of α -Synucleinopathy brains.

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Accumulated α -syn is regarded as a key feature of α -Synucleinopathy. In many neurodegenerative diseases, such as Parkinson's disease (PD), dementia with Lewy bodies (DLB), and multiple system atrophy (MSA), show an α -Synucleinopathy phenotype. Intercellular delivery of α -syn in α -Synucleinopathy occurs through direct penetration, endocytosis, nanotube tunneling-mediated pore formation, and diffusion (Ubhi et al., 2011; Konno et al., 2012; Tosatto et al., 2012; Dieriks et al., 2017; Eguchi et al., 2017). Cell-to-cell delivery of α -syn contributes to neurodegeneration (Desplats et al., 2009; Bruck et al., 2016). Many studies have provided evidence that α -syn transmission occurs in a prion-like manner (Bernis et al., 2015; Steiner et al., 2018). However, this finding remains controversial because of the incomplete understanding of the factors that control the spread of pathogenic proteins and how they work.

Aggregated α -syn leads to many pathological features, such as mitochondrial dysfunction (Liu et al., 2009; Cali et al., 2012; Vicario et al., 2018), dysregulation of calcium homeostasis (Cali et al., 2012; Melachroinou et al., 2013), neuroinflammation, endoplasmic reticulum (ER) stress, protein quality control impairment (Melo et al., 2018; Rocha et al., 2018), Golgi fragmentation (Gosavi et al., 2002), and lysosomal dysfunction (Meredith et al., 2002). In MSA, unlike PD and DLB, aggregated α-syn mainly appears in oligodendrocyte, also called glial cytoplasmic inclusions (GCIs; Ahmed et al., 2013; Bruck et al., 2016; Mori et al., 2020). Gila cells are nonneuronal cells in brain and play a critical role in maintaining neuronal system. The majority of brain cells are glial cells, and they modulate neurogenesis and synaptogenesis (Argente-Arizon et al., 2017). Moreover, glial cells affect brain-blood barrier (BBB) development and function through interactions with neurons and endothelial cells to defend the brain from pathogens (Banerjee and Bhat, 2007; Broux et al., 2015). Glia cells are composed of astrocytes, microglia, and oligodendrocytes in the CNS (Bruck et al., 2016). A major function of astrocytes and microglia involve the immune response. Under pathological conditions, microglia and astrocytes are activated. Activated microglia and astrocytes can release pro-inflammatory cytokines such as interleukin-1 (IL-1), tumor necrosis factor alpha (TNF- α), and interleukin-6 (IL-6; Liu et al., 2011; Tjalkens et al., 2017). These cytokines lead to the production of reactive oxidative stress (ROS) and dysfunction of the BBB. Finally these factors induce neuronal cell death (Yang et al., 2007; Pan et al., 2011) and cause neurodegenerative disease. In general, protein aggregation in affected neurons is also detected in glial cells (Li and Haney, 2020). In particular, α-syn inclusions in glial cells causes a reduction in trophic support, which in turn leads to neuronal loss (Bruck et al., 2016).

Protein aggregation generally occurs in neurodegenerative diseases. In glial cell, aggregated proteins have also been observed (Li and Haney, 2020). Currently, many studies emphasize the role of mitochondria in glia (Yang et al., 2017; McAvoy and Kawamata, 2019; Yan et al., 2020). The mitochondria of glial cells regulate calcium homeostasis, ATP production and the inflammatory response (Yang et al., 2017). Mitochondrial impairment in glia can affect neuronal survival, so glial mitochondrial dysfunction

has recently emerged as a major etiology of neurodegeneration (Yang et al., 2017; McAvoy and Kawamata, 2019; Yan et al., 2020).

In this review, we provide new insights into the roles of glial cells in α -syn-related neurodegenerative diseases and examine the importance of glial mitochondria in the disease progression of α -Synucleinopathy.

NEURODEGENERATIVE DISORDERS ASSOCIATED WITH α-SYNUCLEINOPATHIES

PD

Parkinson's disease is the second most common neurodegenerative disease and is defined as a α -Synucleinopathy. The main pathological features of PD are the selective loss of dopaminergic neurons in the substantia nigra pars compacta (SNpc; Damier et al., 1999; Kalia and Lang, 2015) and protein aggregates (called Lewy bodies and Lewy neurites) consisting mainly of α -syn proteins present in neurons (Spillantini et al., 1997; Gomez-Benito et al., 2020). Moreover, PD has clinical features such as cognitive impairment and motor dysfunction including stiffness, postural instability, and akinesia. PD is mostly known as a sporadic disease, but several gene mutations, such as α-syn, LRRK2, PINK1, DJ-1, and parkin, cause disease (Davie, 2008). Moreover, mutations in α -syn (duplications, triplications, or point mutations) facilitate aggregate formation in neurons and affect the exacerbation and progression of PD. Many pathological features of PD, such as mitochondrial dysfunction (Park et al., 2018; Wang X.L. et al., 2020), neuroinflammation (Wang et al., 2015; Pajares et al., 2020), ER stress (Colla, 2019), and impaired protein quality control (Cook and Petrucelli, 2009; Sala et al., 2016), can lead to neuronal toxicity. α-Synucleinopathy features the accumulation of aggregated α -syn in neuronal and glial cells (McCann et al., 2014). Previous studies showed that healthy neurons transplanted into the striatum of PD patient exhibited α -syn pathology (Kordower et al., 2008; Li et al., 2008). These results indicated that α -syn can propagate into other cells. Moreover, cell-to-cell transmission of aggregated a-syn accelerates Synucleinopathy. Mitochondrial dysfunction is a key feature in PD progression, which is caused by disrupted mitochondrial respiration, decreased mitochondrial membrane potential, and impaired mitophagy (Lin et al., 2019). Moreover, α-syn also influences mitochondrial function (Devi et al., 2008; Pozo Devoto and Falzone, 2017). α-Synucleinopathy induces mitochondrial dysfunction, especially in the context of calcium homeostasis (Vicario et al., 2018; Lin et al., 2019). Mitochondria regulate calcium levels through interactions with ER. However, abnormal α -syn accumulates in both the mitochondria and ER and stresses the ER to release too much calcium into mitochondria. Excessive calcium in mitochondria induces ROS, which can lead to cell death (Melo et al., 2018; Lin et al., 2019). In addition, α-syn is associated with another mitochondrial dynamics, such as the regulation of morphology (fission and fusion), axonal transport and mitophagy (Pozo Devoto and Falzone, 2017).

DLB (Dementia With Lewy Bodies)

Lewy body dementia (LBD) consists of DLB and Parkinson's disease dementia (PDD), and it shares pathological characteristics with PD, such as the accumulation of α -syn, leading to neuronal loss (Kim et al., 2014; Gomperts, 2016; Surendranathan et al., 2020). These patients commonly show parkinsonism motor symptoms, neuropsychiatric symptoms, cognitive defects, sleep disorders, and visual hallucinations (McKeith et al., 2017; Velayudhan et al., 2017). Moreover, depressive symptom is also common in DLB patients. Several studies indicated that depression is associated with AD and DLB which cause cognitive and memory dysfunction (Fritze et al., 2011; Rapp et al., 2011). However, the frequency and severity of depression are more severe DLB than AD (Fritze et al., 2011; Chiu et al., 2017). Although LBD and PD have some common features, such as α -syn aggregates and clinical symptoms, they are distinguished by the relative onset and prognosis. DLB is diagnosed as dementia within 1 year after parkinsonian onset. On the other hand, PDD is diagnosed with dementia 1 year after parkinsonian onset. PDD patients show severe movement dysfunctions, whereas DLB patients show severe cognitive impairments (Hansen et al., 2019). DLB is caused by the abnormal accumulation of α-syn in neurons, called Lewy bodies (LBs) and Lewy neurites (LNs; Cummings, 2004; Walker et al., 2015). Most DLB cases occur sporadically, but several papers have suggested that DLB has a genetic cause, such as APOE, SNCA, and LRRK2 (Walker et al., 2015; Orme et al., 2018). Excessive accumulation of α -syn in DLB is associated with the loss and dysfunction of dopaminergic neurons and cholinergic neurons (Tiraboschi et al., 2000; Gomperts, 2016). Cholinergic neurons participate in memory function and age-related dementia including Alzheimer's disease (Haam and Yakel, 2017). In addition to LB and LN pathology, most DLB patients also have amyloid plaque and neurofibrillary pathology in their brains (Perry et al., 1993; Bohnen and Albin, 2011). Unfortunately, the clinical diagnosis of DLB remains difficult. Because the symptoms of DLB are similar to those of other diseases, such as AD, PD, and PDD (Tousi, 2017), DLB symptoms can overlap with those of AD and PDD at the same time (McKeith et al., 2017). Therefore, DLB is usually diagnosed after death.

MSA (Multiple System Atrophy)

Multiple system atrophy is an uncommon neurodegenerative disease that is pathologically characterized by a combination of parkinsonism, cerebellar ataxia, and autonomic dysfunction (Lopez-Cuina et al., 2018; Monzio Compagnoni and Di Fonzo, 2019). MSA is divided into two subtypes, MSA with parkinsonism features (MSA-P), which exhibits bradykinesia, rigidity, postural instability, and tremor, or MSA with cerebellar features (MSA-C), which exhibits gait ataxia, ataxic dysarthria, limb ataxia, and sustained gaze-evoked nystagmus (Gilman et al., 1999). MSA is caused by the aggregation of α -syn in oligodendrocytes, which are called GCIs and are regarded as a key feature of this disease (Asi et al., 2014; Djelloul et al., 2015; Monzio Compagnoni and Di Fonzo, 2019). Moreover, one case report suggested that α -syn co-localize with phosphorylated tau in GCIs in MSA

patients (Piao et al., 2001). Normally, oligodendrocytes exhibit very low-level expression of α -syn (Miller et al., 2005). The mechanisms of a-syn accumulation in oligodendrocytes are not fully understood. Recently, some researchers suggested that the interaction between neurons and oligodendrocytes affects GCIs. The overexpression of α -syn in neurons induces extracellular secretion of α -syn. Then, oligodendrocytes absorb α -syn into the cell in clathrin-dependent manners. In addition, intracellular aggregated α -syn in neurons also increases α -syn release from neurons and this process is mediated by vesicle trafficking (Kisos et al., 2012; Konno et al., 2012; Monzio Compagnoni and Di Fonzo, 2019). Originally, prion-like α-syn propagation was examined in PD (Chu and Kordower, 2015). Monomeric α -syn aggregates into β -sheet structure, which are important features of the self-propagation of fibrillar α -syn aggregates (Chu and Kordower, 2015; Woerman et al., 2018). There are several pathological features of MSA, such as inflammation (Ishizawa et al., 2004; Jellinger, 2014) and impairments in protein degradation (Schwarz et al., 2012; Tanji et al., 2013). Moreover, mitochondrial impairment is implicated in MSA pathogenesis. A previous study showed that the coenzyme Q2 (COQ2) mutation induces MSA. The COQ2 mutation inhibits the synthesis of COQ10, which located in the mitochondrial inner membrane. COQ10 plays a role in the mitochondrial respiratory chain. Thus, loss-of-function of COQ10 increases oxidative stress and induces apoptotic cell death (Quinzii and Hirano, 2010; Multiple-System Atrophy Research Collaboration, 2013). It is already known that oxidative stress is a major contributor to the loss of oligodendrocytes (Bradl and Lassmann, 2010).

PAF (Pure Autonomic Failure)

Unlike PD, DLB, and MSA, Pure autonomic failure (PAF) is a rare sporadic neurodegenerative disease induced by dysfunction of autonomic nervous system (Thaisetthawatkul, 2016). It is slowly progressive disease and caused by abnormal accumulation of a-syn (Thaisetthawatkul, 2016; Coon et al., 2019; Coon, 2020). PAF patients showed orthostatic hypotension, syncope, falls, sexual and bladder dysfunction, and constipation with no motor dysfunction (Stubendorff et al., 2012; Allan, 2019). The pathological characteristic of PAF is the degeneration of peripheral autonomic neurons which have a-syn aggregates (Singer et al., 2017). Furthermore, accumulation of cytoplasmic α -syn inclusion in brainstem nuclei contributes to PAF (Garland et al., 2013; Coon et al., 2019). Donadio et al. (2013) suggested that α -syn aggregation founded in sympathetic nerve fiber using skin biopsy, it might be a biomarker for PAF diagnosis. However, α-syn induced autonomic dysfunction is not clearly understood and there is no cure for PAF (Thaisetthawatkul, 2016). Recent studies suggested that abnormal α -syn in autonomic neurons of PAF patients is able to propagate into central nerve. Wang X.L. et al. (2020, Wang X. J. et al. 2020) reported that injection of α -syn preformed fibrils (PFF) into stellate and celiac ganglia in mutant form α -syn-expressing mice produces propagation of α -syn throughout autonomic pathway into CNS. They also showed that the PFF injected mice have PAF-related pathology. Moreover, about 70% of PAF patients showed rapid eye movement sleep behavior disorder (RBD) strongly associated with the function of



the CNS ("Consensus statement on the definition of orthostatic hypotension, PAF, and MSA. The Consensus Committee of the American Autonomic Society and the American Academy of Neurology" No authors listed, 1996; Coon et al., 2019). Besides, more than 10% of patients initially diagnosed with PAF developed CNS diseases such as PD, DLB, and MSA (Muppidi and Miglis, 2017; Singer et al., 2017). Converting ratio of PAF to CNS diseases is twice as higher MSA than PD or DLB, and it might be associated with low circulating level of norepinephrine in the plasma of MSA and PAF patients (Goldstein et al., 1989).

The accumulation of α -syn aggregates might be the major cause of neurodegeneration in α -Synucleinopathy (**Figure 1**). Thus, there are several α -syn-mediated therapeutic approaches, such as prevention of α -syn aggregates, increasing of α -syn degradation and α -syn immunotherapy, in all α -syn-mediated diseases (Brundin et al., 2017).

α-SYNUCLEIN-INDUCED PATHOLOGICAL CHANGES IN GLIAL CELLS

Oligodendroglia (Oligodendrocytes)

Oligodendrocytes, together with astrocyte and microglia, are glial cells in CNS (Jakel and Dimou, 2017). Oligodendrocytes are myelin-forming cells in the CNS, accounting for approximately 5–8% of all cells in the adult brain (Levine et al., 2001).

Myelin is essential for the transmission of electrical signals along the axon and additionally plays a role in supporting neuronal activity by regulating axon and neuro homeostasis through the supply of neurotrophic factors (Dawson, 2003; Wilkins et al., 2003; Bean, 2007; Peferoen et al., 2014; Kuhn et al., 2019). Demyelination can be caused by damage to oligodendrocytes or the myelin sheaths maintained by these cells (Arenella and Herndon, 1984); moreover, demyelination induces axonal degeneration and oligodendrocyte death, ultimately promoting neuronal death (Bradl and Lassmann, 2010; Kuhn et al., 2019). MS is a demyelination disease caused by inflammation in the CNS. In a previous study, overexpression of wild-type human α-syn in oligodendrocytes induced mitochondrial dysfunction in transgenic mice, leading to oxidative stress in vivo and neuronal cell death (Stefanova et al., 2005). The pathological features of oligodendrocytes are associated with ROS and the inflammatory response (Thorburne and Juurlink, 1996; Jurewicz et al., 2005). Normally, oligodendrocytes cooperate with astrocytes to regulate the immune response. A recent study suggested that nuclear factor κB (NF- κB) is a main inflammatory modulator in MS, and plays a role in myelin formation (Nickols et al., 2003; Stone et al., 2017). NF-κB predominantly localizes in the cytoplasm, and activated NF-KB translocate to the nucleus and activates the transcript of inflammatory genes including cytokines, chemokines, and adhesion genes (Lawrence, 2009). Moreover, another study showed that NF-KB activation in oligodendrocytes could drive experimental autoimmune encephalomyelitis (EAE) and the expression of interferon-y (IFN- γ) to exacerbate the inflammatory response in an *in vivo* model (Lin et al., 2012, 2013; Stone et al., 2017). Additionally, oligodendrocytes contact surrounding nerve cells to provide neurotrophic factors such as glial cell line-derived neurotrophic factor (GDNF), brain-derived neurotrophic factor (BDNF), and insulin-like growth factor 1 (IGF-1) for protection (Bradl and Lassmann, 2010). Oligodendrocytes do not express α -syn under normal conditions (Kahle et al., 2002), but aggregated α -syn was found in oligodendrocytes in midbrains of PD and DLB patients (Wakabayashi et al., 2000). Furthermore, phosphorylated α -syn was observed in GCIs (Fujiwara et al., 2002). Abnormal accumulation of α -syn occurs with the demyelination and brain atrophy associated with neurodegeneration (Yoon et al., 2020). We already mentioned that MSA is caused by aggregated α -syn in GCIs. A recent paper showed that α -syn aggregation was observed in the oligodendrocytes of human MSA-P patients (May et al., 2014).

Several papers have shown that the aggregation of α -syn is accelerated by heparin and heparan sulfate (HS; Cohlberg et al., 2002; Ihse et al., 2017; Maiza et al., 2018). HS is a linear polysaccharide expressed on the cell surface or as an extracellular matrix protein (Iozzo, 1998; Medeiros, 2000). Heparan sulfate proteoglycans (HSPGs) are internalized by α -syn in the MO3.13 human oligodendrocyte cell line (Ihse et al., 2017). Furthermore, the presence of HSPGs initiates a conformational change in α -syn forms a native state into an oligometric state, including β-sheet formation, through protein unfolding (Motamedi-Shad et al., 2009; Maiza et al., 2018). Aggregate α-syn uptake relies on HS by binding to the plasma membrane (Holmes et al., 2013; Ihse et al., 2017). Additionally, several hypotheses of a-syn aggregation exist cell-to-cell propagation of aggregated α -syn from neurons to oligodendrocytes and/or the secretion of α -syn by oligodendrocyte.

Astroglia (Astrocytes)

Astrocytes are the most common type of glial cell in the CNS and account for at least one third of the brain mass. Astrocytes support the physiological function of the neuronal system by regulating blood flow, maintaining the blood-brain barrier and regulating synaptic plasticity (Sofroniew and Vinters, 2010; Chavarria et al., 2018). Moreover, astrocytes are the main cells responsible for glucose metabolism in the brain and release gliotransmitters, including γ -aminobutyric acid (GABA) and glutamate (Lee et al., 2010a). In addition, astrocytes also play major roles in maintaining neuronal homeostasis through the absorption of glutamate and potassium ions (Thorburne and Juurlink, 1996; Simard and Nedergaard, 2004).

A recent study found that astrocytes isolated from neuroinflammatory and ischemic brains showed two reactive phenotypes, A1 and A2 (Liddelow and Barres, 2017). A1 astrocytes are activated by pro-inflammatory microglia, impair synaptic formation, and kill both neurons and oligodendrocytes, while A2 astrocytes appear to produce substances that promote growth and neuronal survival. Although the number of A1 astrocytes are increased in the affected brain regions of many neurodegenerative diseases, including AD, HD, PD, ALS, and MS (Liddelow and Barres, 2017), detailed mechanisms of A1 and A2 induction have not been clearly identified.

In contrast to neurons, astrocytes express α -syn at very low levels. However, cytoplasmic immunoreactivity of a-syn in astrocytes has been clearly observed in the normal human brain (Mori et al., 2002). Some studies suggested that the number of reactive astrocytes did not increase in the SN and putamen regions in PD patients (Mirza et al., 1999; Song et al., 2009). However, other postmortem studies of PD patient brains reported massive increase in reactive astrocytes and the loss of dopaminergic neurons in the SN (Hirsch et al., 2005), while others reported that reactive astrogliosis was mild (Vila et al., 2001). In most postmortem studies using the brains of PD patients (Mythri et al., 2011; Lastres-Becker et al., 2012), increased expression of glial fibrillary acidic protein (GFAP) was used as a marker for reactive astrocytes (Eng et al., 1971; Dahl and Bignami, 1976). However, since a significant level of GFAP is already present in non-reactive astrocytes, it seems difficult to accurately measure the number of reactive astrocytes with GFAP immunostaining (Sofroniew and Vinters, 2010; Rusnakova et al., 2013).

Astrocytes also have phagocytic activity to degrade cellular debris and synaptic elements (Morizawa et al., 2017). Accumulating evidence suggests that the loss of proper phagocytic functions in astrocytes may contribute to neurodegenerative processes (Jones et al., 2013; Tong et al., 2014; Chung et al., 2016). Unfortunately, whether astrocytic receptors are involved in the recognition of α -syn aggregates and/or their delivery to lysosomes is still unknown.

Astrocytes respond to pathological stimuli by secreting inflammatory cytokines and increasing the level of GFAP. However, in the context of PD, it is still not clear whether astrocyte activation plays a positive or negative role in disease progression (Tremblay et al., 2019).

Both microglia and astrocytes can take up monomeric and fibrillar forms of α-syn (Stefanova et al., 2011; Fellner et al., 2013). In microglia, TLR4 has been implicated in α -syn clearance, the secretion of pro-inflammatory cytokines, and the production of ROS (Fellner et al., 2013). However, astrocytes do not require TLR4 for α -syn uptake (Fellner et al., 2013). Interestingly, α -syn oligomers are rapidly internalized by astrocytes, but internalized α -syn is not completely degraded by lysosomes. Thus, α -syn oligomers accumulate intracellularly and this accumulated α -syn induces mitochondrial defects (Lindstrom et al., 2017). Another study has also reported that the accumulation of α -syn in human astrocytes affects their phagocytic-lysosomal mechanism and that aggregated α -syn is transported through newly formed tunneling nanotubes (Rostami et al., 2017). Evidence suggests that astrocytic a-syn accumulation is implicated in neurotoxic alterations in astrocytes, but the underlying mechanism is still not clear.

Microglia

Although microglia account for the smallest proportion (10% or less) of healthy mammalian brain glia (von Bartheld et al., 2016), they play an important role in the immune response in the CNS. Microglia also regulate neuronal activity and viability in the adult brain through direct contact with neurons (Li et al.,

2012; Kohman et al., 2013) and the secretion of soluble factors (Ramirez et al., 2005; Tichauer et al., 2007).

The morphology and transformative ability of microglia are similar to those of peripheral macrophages/monocytes (Kreutzberg, 1996). The phagocytic activity of microglia is important for the developmental process, as well as injury repair in the adult brain (Neumann et al., 2009).

During brain development, a significant proportion of cells must be eliminated by programmed cell death, while the remaining cells differentiate and migrate. Microglia engulf dving cells, and this phagocytic activity contributes to the proper patterning of the developing brain (Mariin-Teva et al., 2004; Frost and Schafer, 2016). In addition, microglia remove excess synapses in the developing brain and appear to play an important role in the wiring of the CNS (Jakel and Dimou, 2017). Under pathological conditions such as neurodegenerative diseases, infection and physical injury, microglia are activated. Activated microglia are polarized into two different states, M1 (pro-inflammatory), and M2 (anti-inflammatory), according to the signals in the surrounding environment and function in immune regulation (Moehle and West, 2015). M1 microglia release several pro-inflammatory cytokines, including TNF- α , IL-6, IL-12, and IL-1 β , followed by CCL2 and CXCL10. In contrast to M1 cells, M2 microglia are associated with anti-inflammatory functions and promote wound healing and tissue repair. M2 microglia secrete major anti-inflammatory cytokines, such as IL-4, IL-13, IL-10, and TGF-B (Du et al., 2018). In PD, affected DA neurons in the brains of PD patients are surrounded by M1 microglia (Tang and Le, 2016). Moreover, microglia-mediated neuroinflammation shows an inverse relationship to the survival of dopaminergic neurons in PD patients (Tang and Le, 2016). Increased microglial activation has been predominantly found in the substantia nigra (SN) and other brain regions associated with pathological a-syn accumulation (Doorn et al., 2014). In addition, overexpression of the mutant a-syn in microglia induces polarization to the M1 phenotype, which is characterized by increased levels of pro-inflammatory cytokines (such as TNF- α and NO; Rojanathammanee et al., 2011). The role of M2 microglia in PD is not well understood, but the shift from the M1 proinflammatory state to the anti-inflammatory M2 state might be more beneficial in neuroprotection than simply blocking microglial activation (Subramaniam and Federoff, 2017). Thus, a therapeutic approach promoting M1 to M2 polarization could be promising in PD.

Toll-like receptors (TLRs) are known to be involved in the innate immune response, cell survival and death. TLRs are expressed in neurons (Tang et al., 2007), astrocytes (Bowman et al., 2003), microglia (Olson and Miller, 2004), and oligodendrocytes (Lehnardt et al., 2002) in the CNS. Many studies have reported an association between TLRs, especially TLR2, 4 and 9, and neurodegenerative diseases such as PD, AD and ALS. Several studies have shown that increased expression of various receptors, including TLR2 (Kim et al., 2013), TLR4 (Fellner et al., 2013), cluster of differentiation (CD) 11b (Hou et al., 2018), and CD36, (Su et al., 2008) by α -syn aggregation can lead to receptor-mediated activation of the inflammatory signaling cascade. TLR2 is one of the most well-studied TLRs in the context of neurodegenerative diseases. In mice, overexpression of α -syn increased TLR2 expression and microglial activation (Drouin-Ouellet et al., 2014).

The α -syn released by neurons acts as an endogenous ligand for TLR2 on microglia. TLR2- α -syn binding activates the inflammatory responses of microglia, which eventually produce toxic molecules that cause neurodegeneration (Kim et al., 2013, 2016). Previous studies have shown that TLR4 also plays an important role in α -Synucleinopathies.

Stimulation of TLR4 by monophosphoryl lipid A, a selective TLR4 agonist, in an α -syn-overexpressing mouse model, increased α -syn clearance by microglia and suppressed disease-like phenotype, such as motor deficiency (Venezia et al., 2017). In BV2 microglia-like cells, the TLR4-dependent immune response is modulated by Nurr1 and NF- κ B signaling, and this TLR4-mediated neuroinflammation pathway is mediated by α -synuclein (Shao et al., 2019). In addition, Choi et al. (2020) showed that TLR4-NF- κ B-p62-mediated selective autophagy activation in microglia was implicated in the clearance of neuron-secreted α -syn and this clearance mechanism was important for neuroprotective functions of microglia.

GLIAL MITOCHONDRIAL DYSFUNCTION IN α -SYNUCLEINOPATHIES

Cell Death

Severe mitochondrial damage can induce cell death via necrosis or apoptosis (Cillero-Pastor et al., 2011). Several studies have shown that the interaction between α -syn and cytochrome C oxidase (COX, mitochondrial complex IV) induces both α -syn aggregation and mitochondrial dysfunction, causing neurodegeneration (Irrinki et al., 2011; Al-Mansoori et al., 2013; Ciccone et al., 2013).

 α -syn acts as a regulator of mitochondrial homeostasis, and mitochondrial deficiency and impairment are a key pathological features of PD (Gao et al., 2017; Pozo Devoto and Falzone, 2017). Complex 1 deficiency or dysfunction associated with mitochondrial dysfunction was found in the SN region of PD patients (Schapira et al., 1989; Parker et al., 2008). Furthermore, many *a*-Synucleinopathy models have shown that the level of α -syn expression is associated with cell death and mitochondrial deficiency (Hsu et al., 2000; Bellucci et al., 2008; Trancikova et al., 2012). Numerous studies have revealed that the interactions between mitochondria and α -syn play important roles in both physiological and pathological conditions, but most of these studies focused on their role in neurons (Faustini et al., 2017). However, some recent studies have suggested that α -syninduced mitochondrial defects in glia are also implicated in various pathogenic features, such as neurodegeneration and neuroinflammation.

Accumulating evidence suggests that astrocytes might be a major contributor to the progression of α -Synucleinopathies. For example, α -syn overexpression in astrocytes induces astrocyte activation, and these reactive astrocytes cause neuronal death (Chavarria et al., 2018) and susceptibility to oxidative stress

(Stefanova et al., 2001). Chavarria et al. (2018) investigated the effects of the different species of α -syn (monomer, oligomer, and fibrillar) protein on primary rat cortical astrocyte. All of α -syn species can activate astrocytes, and astrocytes treated with the different a-syn species induced cell death of hippocampal neuron in neuron-astrocyte co-culture. In particular, only oligomeric α-syn induce mitochondrial dysfunction and increased hydrogen peroxided generation in astrocytes, whereas fibrillar α-syn treated astrocytes enhanced the secretion of pro-inflammatory cytokines. Stefanova et al. (2001) showed that overexpression of wildtype α -syn or C-terminal truncated α -syn in U373 astrocytoma cells induced apoptotic cell death and increased susceptibility to oxidative stress, and that it was partially present in the cytoplasmic inclusion. In addition, in vitro and in vivo model studies have demonstrated that α -syn can spread from neurons to glial cells either through secretion to the extracellular space or direct cell-to-cell transfer (Angot et al., 2012; Lee et al., 2012; Reves et al., 2015). α -syn inclusions are mainly found in neurons, but also frequently detected in glial cells. Lindstrom et al. (2017) tried to elucidate the clearance mechanism of toxic α -syn in glial cells. This study examined the uptake, degradation, and toxic effects of oligometric α -syn in the co-culture system of primary mouse neurons, astrocytes, and oligodendrocytes. Astrocytes rapidly and extensively absorb α -syn from the extracellular space and sequester it. In the early stage of α -Synucleinopathy, this sequestration has been shown to play a role in preventing toxicity and disease progression (Lindstrom et al., 2017; Lin et al., 2019). Astrocytes degrade absorbed α -syn oligomer via the autophagy lysosomal degradation pathway, but remaining intracellular deposition of a-syn causes mitochondrial dysfunction and cell death (Lindstrom et al., 2017). Another study showed that overexpression of wild-type and disease associated mutant (A30P and A53T) a-syn in immortalized astrocytes decreased LC3-II and increased p62 protein levels, suggesting the suppression of autophagy. In addition, mitochondrial membrane potential loss and increased parkin expression were evident in mutant α -syn expressing astrocytes (Erustes et al., 2018). Moreover, a recent study demonstrated that overexpression of mutant a-syn (A30P and A53T) in primary rat astrocytes induced an unfolded protein response through the PERK/eIF2alpha signaling pathway. In particular, dysfunction of the ER-Golgi compartment in astrocytes overexpressing wild type α -syn can inhibit neurite growth by reducing GDNF levels (Liu et al., 2018). In another study, α-syn was transfected into ONL-t40 cells, an oligodendroglial cell line. This study showed that α -syn overexpression induces mitochondrial dysfunction, impaired autophagic flux, and the formation of a-syn aggregates in ONLt40 cell lines (Pukass et al., 2015). It is clear that mitochondrial impairment is implicated in α-syn-induced glial cell death, but the molecular mechanisms of glial mitochondrial dysfunction and cell death in α -Synucleinopathies are not fully understood.

Inflammation

Mitochondrial dysfunction is associated with the induction of the inflammatory response. Mitochondrial impairment increases mtROS production, extracellular ATP levels and mtDNA release, all of which induce a vicious inflammatory response cycle that exacerbates mitochondrial dysfunction (Lopez-Armada et al., 2013).

Among the various putative factors contributing to PD etiology, the neuroinflammatory mechanism serves as a major contributor. Increased pro-inflammatory cytokines (including IL-1 β , IL-6, TNF, and IFN- γ) have been observed in postmortem brains and cerebrospinal fluid from PD patients (Mogi et al., 1994a,b), moreover, the blood serum of PD patients also showed increased concentration of inflammatory cytokines (including IL-2, IL-6, TNF-α, and IFN-γ; Reale et al., 2009). Neuroinflammation is primarily mediated by the activation of glial cells and is accompanied by the production of inflammatory cytokines. Microglia have traditionally been considered to play a major role in the immune response in the CNS (Soulet and Rivest, 2008). Pathological α -syn aggregation in PD causes microglial activation, which is known to strongly induce microglial neuroinflammatory responses in the brain (Zhang et al., 2005). Microglia that absorb α-syn aggregates via phagocytosis promote NADPH oxidase activation and ROS production, leading to the secretion of pro-inflammatory neurotoxic cytokines (Zhang et al., 2005; Walter and Neumann, 2009) and chemokines (Fellner et al., 2013; Roodveldt et al., 2013). In addition, during the inflammatory response, changes in mitochondrial metabolism contribute to microglial activation.

The levels of inflammatory cytokines, such as IL-6, and the number of activated microglia are increased in the hippocampus in patients with PD and DLB (Imamura et al., 2005). M1related cytokines, such as TNF- α (Mogi et al., 1994b) and IL-6 (Muller et al., 1998), were increased in the cerebrospinal fluid of PD patients, and the levels of these cytokines correlated with poor prognosis. A number of studies have shown that α -syn induces the pro-inflammatory response in microglia or macrophage to produce the inflammatory cytokines such as IL-1 β , IL-6, and TNF- α (Klegeris et al., 2008; Lee et al., 2009). Inflammatory mediators such as iNOS and COX2 were also upregulated in activated microglia by wild-type α -syn treatment. This result suggested that that CD36 scavenger receptor and downstream kinases are involved in α -syn-mediated microglial activation. (Su et al., 2008). Hoenen et al. (2016) also showed that a disease associated α -syn mutant (A35T) more strongly induced reactive microglia than the human wild-type protein in a transgenic mouse model. The researchers also found that the NF-kB/AP-1/Nrf2 pathway was implicated in A35T α-syn-induced microglial activation via MAPK. In addition, Rannikko et al. (2015) reported that overexpression of exogenous wild type α -syn in primary mouse astrocytes can induce inflammatory responses such as induction of NO synthase and COX-2 expression.

Furthermore, in the Fellner (2013; Fellner et al., 2013) study, purified microglia and astrocytes from wild-type (TLR4 +/+) and TLR4-deficient (TLR4-/-) mice were treated with α -syn (full length soluble, fibrillized, and C-terminally truncated), and phagocytic activity, NF- κ B nuclear translocation, cytokine release and ROS production were measured. Treatment of TLR4+/+ glial cells with C-terminally truncated α -syn induced upregulation of phagocytic activity, NF- κ B translocation, ROS production, and the release of IL-6, CXCL1, and TNF- α (Fellner et al., 2013). The stefanova (2011; Stefanova et al., 2011) study reported that α -syn clearance was reduced in TLR4 deficiency murine. Moreover, in an animal model study of MSA based on oligodendroglial α -syn overexpression, TLR4 deficiency increased motor impairment, and increased loss of nigrostriatal dopaminergic neurons. However, the role of TLR4 in α -syndependent activation of microglial and astrocytes has not been clearly elucidated.

To date, many studies have shown that activated astrocytes also play an important role in neuroinflammation in the aged brain and most neurodegenerative diseases. Activated astrocytes release inflammatory cytokines and chemokines that can cause neuronal damage (Komine et al., 2018; Yu et al., 2018). Although the exact pathogenic mechanism of PD is unclear, accumulating evidence suggests that reactive astrogliosis and astrocyte-mediated neuroinflammation play important roles in the pathogenesis of PD (Wang et al., 2015; Liddelow et al., 2017). In another study, inflammation-related factors such as GFAP, COX-2, iNOS, TNF- α , and IL-1 β were increased in primary rat astrocytes by A53T α -syn aggregate treatment via the NF- κ B and c-Jun N-terminal kinase signaling pathways, and the upregulation of heat shock protein 70 effectively suppressed the α -syn-induced neuroinflammatory response (Yu et al., 2018).

Calcium Homeostasis

Calcium is essential for cellular signaling, and calcium homeostasis is important for neuronal integrity because it is involved in neuronal plasticity, synaptic transmission, and cell survival. In neurons, calcium channels are the key signaling elements that regulate the release of biological factors, such as neurotransmitters and hormones, through calcium sensing (Leandrou et al., 2019). Among the causes of α -syn pathology, the breakdown of calcium homeostasis promotes aggregation and abnormal secretion of α -syn (Wojda et al., 2008; Hettiarachchi et al., 2009; Yang et al., 2019). Cell culture model studies suggested that a temporary increase in intracellular calcium concentration induced cytoplasmic α -syn aggregates (Nath et al., 2011; Follett et al., 2013).

Nath et al. (2011) showed that the addition of calcium to the recombinant α -syn monomer *in vitro* triggers oligomer formation in a dose-dependent manner. Besides, in 1321N1 glioma cells expressing α -syn-GFP, when treated with thapsigargin or calcium ionophore to induce a temporary



increase in intracellular free calcium, α -syn aggregates in the cytoplasm were significantly increased. Another study show that raised intracellular Ca²⁺ mediated by K⁺ depolarization can lead to cytoplasmic α -syn aggregation (Follett et al., 2013).

Dufty et al. (2007) demonstrated that cleavage of α -syn by calpain I, a calcium-dependent protease, can facilitate the aggregation of α-syn. Moreover, Luth et al. (2014) investigated the relationship between mitochondrial Ca^{2+} stress and α -syn using isolated mitochondria and purified recombinant human α -syn. Interestingly, the mitotoxic effect of α -syn was particularly dependent on electron flow through complex I and the mitochondrial uptake of exogenous Ca²⁺. Consequently, the soluble prefibrillar α-syn oligomer induces several mitochondrial phenotypes (changes in membrane potential, disruption of Ca²⁺ homeostasis, mitochondrial complex I dysfunction, and enhancement of cytochrome c release) that have been observed in vitro and in vivo models of PD. However, the mechanism by which calcium promotes α -syn aggregation is not clear, and the interaction between calcium and α -syn aggregation in glial cells is not been clearly characterized.

CONCLUSION

Links Between Glial Mitochondrial Dysfunction and Neurodegeneration in α-Synucleinopathies

Many *in vitro* and *in vivo* animal studies have focused on neuron-to-neuron propagation of α -syn aggregates (Lee et al., 2012; Dehay et al., 2016). However, studies on α -syn in the context of neuronal dysfunction and studies on the association of α -syn accumulation and propagation in glial cells have been largely overlooked. Some recent studies have provided evidence for the pathogenic response of glial cells to neuron-derived α -syn aggregates. α -syn secreted from neurons induces mitochondrial impairment in microglia, and α -syn-induced mitochondrial dysfunction can activate the microglia-mediated neuroinflammatory response (**Figure 2**). Pro-inflammatory M1 microglia secrete neurotoxic agents that cause neurodegeneration. In addition, it has been found that the

REFERENCES

- Ahmed, Z., Asi, Y. T., Lees, A. J., Revesz, T., and Holton, J. L. (2013). Identification and quantification of oligodendrocyte precursor cells in multiple system atrophy, progressive supranuclear palsy and Parkinson's disease. *Brain Pathol.* 23, 263–273. doi: 10.1111/j.1750-3639.2012.00637.x
- Allan, L. M. (2019). Diagnosis and management of autonomic dysfunction in dementia syndromes. *Curr. Treat. Options Neurol.* 21:38. doi: 10.1007/s11940-019-0581-2
- Al-Mansoori, K. M., Hasan, M. Y., Al-Hayani, A., and El-Agnaf, O. M. (2013). The role of alpha-synuclein in neurodegenerative diseases: from molecular pathways in disease to therapeutic approaches. *Curr. Alzheimer. Res.* 10, 559–568. doi: 10.2174/1567205011310060002
- Angot, E., Steiner, J. A., Lema Tome, C. M., Ekstrom, P., Mattsson, B., Bjorklund, A., et al. (2012). Alpha-synuclein cell-to-cell transfer and seeding in grafted dopaminergic neurons in vivo. *PLoS One* 7:e39465. doi: 10.1371/journal.pone. 0039465

neurotoxic response of microglia is mediated by the activation of TLR2, a receptor for neuron-derived α -syn (Kim et al., 2013, 2016). Similar to microglia, pathogenic α -syn-treated astrocytes activate pro-inflammatory responses, including the production of pro-inflammatory cytokines and chemokines (Lee et al., 2010b). Although mitochondrial dysfunction in glial cells is one of the major contributors to neuroinflammation, which causes neurodegeneration in α -Synucleinopathies, the molecular mechanism underlying α -syn-induced mitochondrial impairment is not fully understood. Thus, further studies are warranted to elucidate the non-cell autonomous neurotoxic crosstalk between glial cells and neurons, and this crosstalk mechanism may be a promising target for α -Synucleinopathyassociated neurodegenerative disease treatment.

AUTHOR CONTRIBUTIONS

SL, YK, SK, MJ, and Y-MJ provided ideas for the project and participated in data collection. Y-MJ, YK, and H-JK wrote the manuscript. All authors read and approved the final manuscript.

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SUPPLEMENTARY MATERIAL

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- Arenella, L. S., and Herndon, R. M. (1984). Mature oligodendrocytes. Division following experimental demyelination in adult animals. *Arch. Neurol.* 41, 1162– 1165. doi: 10.1001/archneur.1984.04050220060015
- Argente-Arizon, P., Guerra-Cantera, S., Garcia-Segura, L. M., Argente, J., and Chowen, J. A. (2017). Glial cells and energy balance. J. Mol. Endocrinol. 58, R59–R71. doi: 10.1530/JME-16-0182
- Asi, Y. T., Simpson, J. E., Heath, P. R., Wharton, S. B., Lees, A. J., Revesz, T., et al. (2014). Alpha-synuclein mRNA expression in oligodendrocytes in MSA. *Glia* 62, 964–970. doi: 10.1002/glia.22653
- Banerjee, S., and Bhat, M. A. (2007). Neuron-glial interactions in blood-brain barrier formation. Annu. Rev. Neurosci. 30, 235–258. doi: 10.1146/annurev. neuro.30.051606.094345
- Bartels, T., Choi, J. G., and Selkoe, D. J. (2011). alpha-Synuclein occurs physiologically as a helically folded tetramer that resists aggregation. *Nature* 477, 107–110. doi: 10.1038/nature10324
- Bean, B. P. (2007). The action potential in mammalian central neurons. Nat. Rev. Neurosci. 8, 451–465. doi: 10.1038/nrn2148

- Bellucci, A., Collo, G., Sarnico, I., Battistin, L., Missale, C., and Spano, P. (2008). Alpha-synuclein aggregation and cell death triggered by energy deprivation and dopamine overload are counteracted by D2/D3 receptor activation. J. Neurochem. 106, 560–577. doi: 10.1111/j.1471-4159.2008.05 406.x
- Bernis, M. E., Babila, J. T., Breid, S., Wusten, K. A., Wullner, U., and Tamguney, G. (2015). Prion-like propagation of human brain-derived alphasynuclein in transgenic mice expressing human wild-type alpha-synuclein. Acta Neuropathol. Commun. 3:75. doi: 10.1186/s40478-015-0254-7
- Bohnen, N. I., and Albin, R. L. (2011). The cholinergic system and Parkinson disease. *Behav. Brain Res.* 221, 564–573. doi: 10.1016/j.bbr.2009.12.048
- Bowman, C. C., Rasley, A., Tranguch, S. L., and Marriott, I. (2003). Cultured astrocytes express toll-like receptors for bacterial products. *Glia* 43, 281–291. doi: 10.1002/glia.10256
- Bradl, M., and Lassmann, H. (2010). Oligodendrocytes: biology and pathology. Acta Neuropathol. 119, 37–53. doi: 10.1007/s00401-009-0601-5
- Broux, B., Gowing, E., and Prat, A. (2015). Glial regulation of the blood-brain barrier in health and disease. *Semin. Immunopathol.* 37, 577–590. doi: 10.1007/ s00281-015-0516-2
- Bruck, D., Wenning, G. K., Stefanova, N., and Fellner, L. (2016). Glia and alphasynuclein in neurodegeneration: a complex interaction. *Neurobiol. Dis.* 85, 262–274. doi: 10.1016/j.nbd.2015.03.003
- Brundin, P., Dave, K. D., and Kordower, J. H. (2017). Therapeutic approaches to target alpha-synuclein pathology. *Exp. Neurol.* 298, 225–235. doi: 10.1016/ j.expneurol.2017.10.003
- Burre, J., Sharma, M., and Sudhof, T. C. (2014). Alpha-Synuclein assembles into higher-order multimers upon membrane binding to promote SNARE complex formation. *Proc. Natl. Acad. Sci. U.S.A.* 111, E4274–E4283. doi: 10.1073/pnas. 1416598111
- Cali, T., Ottolini, D., Negro, A., and Brini, M. (2012). alpha-Synuclein controls mitochondrial calcium homeostasis by enhancing endoplasmic reticulummitochondria interactions. *J. Biol. Chem.* 287, 17914–17929. doi: 10.1074/jbc. M111.302794
- Chavarria, C., Rodriguez-Bottero, S., Quijano, C., Cassina, P., and Souza, J. M. (2018). Impact of monomeric, oligomeric and fibrillar alpha-synuclein on astrocyte reactivity and toxicity to neurons. *Biochem. J.* 475, 3153–3169. doi: 10.1042/BCJ20180297
- Chiu, P. Y., Wang, C. W., Tsai, C. T., Li, S. H., Lin, C. L., and Lai, T. J. (2017). Depression in dementia with Lewy bodies: a comparison with Alzheimer's disease. *PLoS One* 12:e0179399. doi: 10.1371/journal.pone.0179399
- Choi, I., Zhang, Y., Seegobin, S. P., Pruvost, M., Wang, Q., Purtell, K., et al. (2020). Microglia clear neuron-released alpha-synuclein via selective autophagy and prevent neurodegeneration. *Nat. Commun.* 11:1386. doi: 10.1038/s41467-020-15119-w
- Chu, Y., and Kordower, J. H. (2015). The prion hypothesis of Parkinson's disease. *Curr. Neurol. Neurosci. Rep.* 15:28. doi: 10.1007/s11910-015-0549-x
- Chung, W. S., Verghese, P. B., Chakraborty, C., Joung, J., Hyman, B. T., Ulrich, J. D., et al. (2016). Novel allele-dependent role for APOE in controlling the rate of synapse pruning by astrocytes. *Proc. Natl. Acad. Sci. U.S.A.* 113, 10186–10191. doi: 10.1073/pnas.1609896113
- Ciccone, S., Maiani, E., Bellusci, G., Diederich, M., and Gonfloni, S. (2013). Parkinson's disease: a complex interplay of mitochondrial DNA alterations and oxidative stress. *Int. J. Mol. Sci.* 14, 2388–2409. doi: 10.3390/ijms14022388
- Cillero-Pastor, B., Martin, M. A., Arenas, J., Lopez-Armada, M. J., and Blanco, F. J. (2011). Effect of nitric oxide on mitochondrial activity of human synovial cells. *BMC Musculoskelet. Disord.* 12:42. doi: 10.1186/1471-2474-12-42
- Cohlberg, J. A., Li, J., Uversky, V. N., and Fink, A. L. (2002). Heparin and other glycosaminoglycans stimulate the formation of amyloid fibrils from alphasynuclein in vitro. *Biochemistry* 41, 1502–1511. doi: 10.1021/bi011711s
- Colla, E. (2019). Linking the endoplasmic reticulum to Parkinson's disease and alpha-synucleinopathy. Front. Neurosci. 13:560. doi: 10.3389/fnins.2019.00560
- Cook, C., and Petrucelli, L. (2009). A critical evaluation of the ubiquitinproteasome system in Parkinson's disease. *Biochim. Biophys. Acta* 1792, 664– 675. doi: 10.1016/j.bbadis.2009.01.012
- Coon, E. A. (2020). Autonomic dysfunction in the synucleinopathies. Semin. Neurol. 10:363. doi: 10.1055/s-0040-1713844
- Coon, E. A., Singer, W., and Low, P. A. (2019). Pure autonomic failure. *Mayo Clin. Proc.* 94, 2087–2098. doi: 10.1016/j.mayocp.2019.03.009

- Cummings, J. L. (2004). Dementia with lewy bodies: molecular pathogenesis and implications for classification. J. Geriatr. Psychiatry Neurol. 17, 112–119. doi: 10.1177/0891988704267473
- Dahl, D., and Bignami, A. (1976). Immunogenic properties of the glial fibrillary acidic protein. Brain Res. 116, 150–157. doi: 10.1016/0006-8993(76)90257-2
- Damier, P., Hirsch, E. C., Agid, Y., and Graybiel, A. M. (1999). The substantia nigra of the human brain. II. Patterns of loss of dopamine-containing neurons in Parkinson's disease. *Brain* 122(Pt 8), 1437–1448. doi: 10.1093/brain/122.8.1437
- Davie, C. A. (2008). A review of Parkinson's disease. *Br. Med. Bull.* 86, 109–127. doi: 10.1093/bmb/ldn013
- Dawson, M. (2003). NG2-expressing glial progenitor cells: an abundant and widespread population of cycling cells in the adult rat CNS. *Mol. Cell. Neurosci.* 24, 476–488. doi: 10.1016/s1044-7431(03)00210-0
- Dehay, B., Vila, M., Bezard, E., Brundin, P., and Kordower, J. H. (2016). Alphasynuclein propagation: new insights from animal models. *Mov. Disord.* 31, 161–168. doi: 10.1002/mds.26370
- Desplats, P., Lee, H. J., Bae, E. J., Patrick, C., Rockenstein, E., Crews, L., et al. (2009). Inclusion formation and neuronal cell death through neuron-to-neuron transmission of alpha-synuclein. *Proc. Natl. Acad. Sci. U.S.A.* 106, 13010–13015. doi: 10.1073/pnas.0903691106
- Dettmer, U., Newman, A. J., Luth, E. S., Bartels, T., and Selkoe, D. (2013). In vivo cross-linking reveals principally oligomeric forms of alpha-synuclein and betasynuclein in neurons and non-neural cells. *J. Biol. Chem.* 288, 6371–6385. doi: 10.1074/jbc.M112.403311
- Dettmer, U., Newman, A. J., von Saucken, V. E., Bartels, T., and Selkoe, D. (2015). KTKEGV repeat motifs are key mediators of normal alpha-synuclein tetramerization: their mutation causes excess monomers and neurotoxicity. *Proc. Natl. Acad. Sci. U.S.A.* 112, 9596–9601. doi: 10.1073/pnas.1505953112
- Devi, L., Raghavendran, V., Prabhu, B. M., Avadhani, N. G., and Anandatheerthavarada, H. K. (2008). Mitochondrial import and accumulation of alpha-synuclein impair complex I in human dopaminergic neuronal cultures and Parkinson disease brain. J. Biol. Chem. 283, 9089–9100. doi: 10.1074/jbc.M710012200
- Dieriks, B. V., Park, T. I., Fourie, C., Faull, R. L., Dragunow, M., and Curtis, M. A. (2017). alpha-synuclein transfer through tunneling nanotubes occurs in SH-SY5Y cells and primary brain pericytes from Parkinson's disease patients. *Sci. Rep.* 7:42984. doi: 10.1038/srep42984
- Djelloul, M., Holmqvist, S., Boza-Serrano, A., Azevedo, C., Yeung, M. S., Goldwurm, S., et al. (2015). Alpha-synuclein expression in the oligodendrocyte lineage: an in vitro and in vivo study using rodent and human models. *Stem Cell Rep.* 5, 174–184. doi: 10.1016/j.stemcr.2015.07.002
- Donadio, V., Incensi, A., Cortelli, P., Giannoccaro, M. P., Jaber, M. A., Baruzzi, A., et al. (2013). Skin sympathetic fiber alpha-synuclein deposits: a potential biomarker for pure autonomic failure. *Neurology* 80, 725–732. doi: 10.1212/ WNL.0b013e3182825127
- Doorn, K. J., Moors, T., Drukarch, B., van de Berg, W. D. J., Lucassen, P. J., and van Dam, A. M. (2014). Microglial phenotypes and toll-like receptor 2 in the substantia nigra and hippocampus of incidental Lewy body disease cases and Parkinson's disease patients. *Acta Neuropathol. Commun.* 2:90. doi: 10.1186/ s40478-014-0090-1
- Drouin-Ouellet, J., St-Amour, I., Saint-Pierre, M., Lamontagne-Proulx, J., Kriz, J., Barker, R. A., et al. (2014). Toll-like receptor expression in the blood and brain of patients and a mouse model of Parkinson's disease. *Int. J. Neuropsychopharmacol.* 18:pyu103. doi: 10.1093/ijnp/pyu103
- Du, R. H., Sun, H. B., Hu, Z. L., Lu, M., Ding, J. H., and Hu, G. (2018). Kir6.1/K-ATP channel modulates microglia phenotypes: implication in Parkinson's disease. *Cell Death Dis.* 9:404. doi: 10.1038/s41419-018-0437-9
- Dufty, B. M., Warner, L. R., Hou, S. T., Jiang, S. X., Gomez-Isla, T., Leenhouts, K. M., et al. (2007). Calpain-cleavage of alpha-synuclein: connecting proteolytic processing to disease-linked aggregation. Am. J. Pathol. 170, 1725–1738. doi: 10.2353/ajpath.2007.061232
- Eguchi, K., Taoufiq, Z., Thorn-Seshold, O., Trauner, D., Hasegawa, M., and Takahashi, T. (2017). Wild-type monomeric alpha-synuclein can impair vesicle endocytosis and synaptic fidelity via tubulin polymerization at the calyx of held. *J. Neurosci.* 37, 6043–6052. doi: 10.1523/JNEUROSCI.0179-17.2017
- Eng, L. F., Vanderhaeghen, J. J., Bignami, A., and Gerstl, B. (1971). An acidic protein isolated from fibrous astrocytes. *Brain Res.* 28, 351–354. doi: 10.1016/ 0006-8993(71)90668-8

- Erustes, A. G., Stefani, F. Y., Terashima, J. Y., Stilhano, R. S., Monteforte, P. T., da Silva Pereira, G. J., et al. (2018). Overexpression of alpha-synuclein in an astrocyte cell line promotes autophagy inhibition and apoptosis. *J. Neurosci. Res.* 96, 160–171. doi: 10.1002/jnr.24092
- Faustini, G., Bono, F., Valerio, A., Pizzi, M., Spano, P., and Bellucci, A. (2017). Mitochondria and alpha-Synuclein: friends or foes in the pathogenesis of Parkinson's disease?. *Genes* 8:377. doi: 10.3390/genes8120377
- Fellner, L., Irschick, R., Schanda, K., Reindl, M., Klimaschewski, L., Poewe, W., et al. (2013). Toll-like receptor 4 is required for alpha-synuclein dependent activation of microglia and astroglia. *Glia* 61, 349–360. doi: 10.1002/glia.22437
- Follett, J., Darlow, B., Wong, M. B., Goodwin, J., and Pountney, D. L. (2013). Potassium depolarization and raised calcium induces alpha-synuclein aggregates. *Neurotox Res.* 23, 378–392. doi: 10.1007/s12640-012-9366-z
- Fortin, D. L., Troyer, M. D., Nakamura, K., Kubo, S., Anthony, M. D., and Edwards, R. H. (2004). Lipid rafts mediate the synaptic localization of alpha-synuclein. *J. Neurosci.* 24, 6715–6723. doi: 10.1523/JNEUROSCI.1594-04.2004
- Fritze, F., Ehrt, U., Hortobagyi, T., Ballard, C., and Aarsland, D. (2011). Depressive symptoms in Alzheimers disease and lewy body dementia: a one-year follow-up study. *Dement. Geriatr. Cogn. Disord.* 32, 143–149. doi: 10.1159/000332016
- Frost, J. L., and Schafer, D. P. (2016). Microglia: architects of the developing nervous system. *Trends Cell Biol.* 26, 587–597. doi: 10.1016/j.tcb.2016.02.006
- Fujiwara, H., Hasegawa, M., Dohmae, N., Kawashima, A., Masliah, E., Goldberg, M. S., et al. (2002). alpha-Synuclein is phosphorylated in synucleinopathy lesions. *Nat. Cell. Biol.* 4, 160–164. doi: 10.1038/ncb748
- Gao, F., Yang, J., Wang, D., Li, C., Fu, Y., Wang, H., et al. (2017). Mitophagy in Parkinson's disease: pathogenic and therapeutic implications. *Front. Neurol.* 8:527. doi: 10.3389/fneur.2017.00527
- Garland, E. M., Hooper, W. B., and Robertson, D. (2013). Pure autonomic failure. Handb. Clin. Neurol. 117, 243–257. doi: 10.1016/B978-0-444-53491-0.00020-1
- Gilman, S., Low, P. A., Quinn, N., Albanese, A., Ben-Shlomo, Y., Fowler, C. J., et al. (1999). Consensus statement on the diagnosis of multiple system atrophy. *J. Neurol. Sci.* 163, 94–98. doi: 10.1016/s0022-510x(98)00304-9
- Goldstein, D. S., Polinsky, R. J., Garty, M., Robertson, D., Brown, R. T., Biaggioni, I., et al. (1989). Patterns of plasma levels of catechols in neurogenic orthostatic hypotension. *Ann. Neurol.* 26, 558–563. doi: 10.1002/ana.410260410
- Gomez-Benito, M., Granado, N., Garcia-Sanz, P., Michel, A., Dumoulin, M., and Moratalla, R. (2020). Modeling Parkinson's disease with the alpha-synuclein protein. *Front. Pharmacol.* 11:356. doi: 10.3389/fphar.2020.00356
- Gomperts, S. N. (2016). Lewy body dementias: dementia with lewy bodies and parkinson disease dementia. *Continuum* 22, 435–463. doi: 10.1212/CON. 000000000000309
- Gosavi, N., Lee, H. J., Lee, J. S., Patel, S., and Lee, S. J. (2002). Golgi fragmentation occurs in the cells with prefibrillar alpha-synuclein aggregates and precedes the formation of fibrillar inclusion. J. Biol. Chem. 277, 48984–48992. doi: 10.1074/ jbc.M208194200
- Gurry, T., Ullman, O., Fisher, C. K., Perovic, I., Pochapsky, T., and Stultz, C. M. (2013). The dynamic structure of alpha-synuclein multimers. J. Am. Chem. Soc. 135, 3865–3872. doi: 10.1021/ja310518p
- Haam, J., and Yakel, J. L. (2017). Cholinergic modulation of the hippocampal region and memory function. J. Neurochem. 142(Suppl. 2), 111–121. doi: 10.1111/jnc. 14052
- Hansen, D., Ling, H., Lashley, T., Holton, J. L., and Warner, T. T. (2019). Review: clinical, neuropathological and genetic features of Lewy body dementias. *Neuropathol. Appl. Neurobiol.* 45, 635–654. doi: 10.1111/nan. 12554
- Hettiarachchi, N. T., Parker, A., Dallas, M. L., Pennington, K., Hung, C. C., Pearson, H. A., et al. (2009). alpha-Synuclein modulation of Ca2+ signaling in human neuroblastoma (SH-SY5Y) cells. *J. Neurochem.* 111, 1192–1201. doi: 10.1111/j. 1471-4159.2009.06411.x
- Hirsch, E. C., Hunot, S., and Hartmann, A. (2005). Neuroinflammatory processes in Parkinson's disease. *Parkinsonism Relat. Disord.* 11(Suppl. 1), S9–S15. doi: 10.1016/j.parkreldis.2004.10.013
- Hoenen, C., Gustin, A., Birck, C., Kirchmeyer, M., Beaume, N., Felten, P., et al. (2016). Alpha-synuclein proteins promote pro-inflammatory cascades in microglia: stronger effects of the A53T mutant. *PLoS One* 11:e0162717. doi: 10.1371/journal.pone.0162717
- Holmes, B. B., DeVos, S. L., Kfoury, N., Li, M., Jacks, R., Yanamandra, K., et al. (2013). Heparan sulfate proteoglycans mediate internalization and propagation

of specific proteopathic seeds. Proc. Natl. Acad. Sci. U.S.A. 110, E3138–E3147. doi: 10.1073/pnas.1301440110

- Hou, L., Bao, X., Zang, C., Yang, H., Sun, F., Che, Y., et al. (2018). Integrin CD11b mediates alpha-synuclein-induced activation of NADPH oxidase through a Rho-dependent pathway. *Redox. Biol.* 14, 600–608. doi: 10.1016/j.redox.2017. 11.010
- Hsu, L. J., Sagara, Y., Arroyo, A., Rockenstein, E., Sisk, A., Mallory, M., et al. (2000). α-synuclein promotes mitochondrial deficit and oxidative stress. Am. J. Pathol. 157, 401–410. doi: 10.1016/s0002-9440(10)64553-1
- Huang, M., Wang, B., Li, X., Fu, C., Wang, C., and Kang, X. (2019). alphasynuclein: a multifunctional player in exocytosis, endocytosis, and vesicle recycling. *Front. Neurosci.* 13:28. doi: 10.3389/fnins.2019.00028
- Ihse, E., Yamakado, H., van Wijk, X. M., Lawrence, R., Esko, J. D., and Masliah, E. (2017). Cellular internalization of alpha-synuclein aggregates by cell surface heparan sulfate depends on aggregate conformation and cell type. *Sci. Rep.* 7:9008. doi: 10.1038/s41598-017-08720-5
- Imamura, K., Hishikawa, N., Ono, K., Suzuki, H., Sawada, M., Nagatsu, T., et al. (2005). Cytokine production of activated microglia and decrease in neurotrophic factors of neurons in the hippocampus of Lewy body disease brains. Acta Neuropathol. 109, 141–150. doi: 10.1007/s00401-004-0919-v
- Iozzo, R. V. (1998). Matrix proteoglycans: from molecular design to cellular function. Annu. Rev. Biochem. 67, 609–652. doi: 10.1146/annurev.biochem.67. 1.609
- Irrinki, K. M., Mallilankaraman, K., Thapa, R. J., Chandramoorthy, H. C., Smith, F. J., Jog, N. R., et al. (2011). Requirement of FADD, NEMO, and BAX/BAK for aberrant mitochondrial function in tumor necrosis factor alpha-induced necrosis. *Mol. Cell. Biol.* 31, 3745–3758. doi: 10.1128/MCB.05303-11
- Ishizawa, K., Komori, T., Sasaki, S., Arai, N., Mizutani, T., and Hirose, T. (2004). Microglial activation parallels system degeneration in multiple system atrophy. *J. Neuropathol. Exp. Neurol.* 63, 43–52. doi: 10.1093/jnen/63.1.43
- Jakel, S., and Dimou, L. (2017). Glial cells and their function in the adult brain: a journey through the history of their ablation. *Front. Cell. Neurosci.* 11:24. doi: 10.3389/fncel.2017.00024
- Jakes, R., Spillantini, M. G., and Goedert, M. (1994). Identification of two distinct synucleins from human brain. *FEBS Lett.* 345, 27–32. doi: 10.1016/0014-5793(94)00395-5
- Jellinger, K. A. (2014). Neuropathology of multiple system atrophy: new thoughts about pathogenesis. Mov. Disord. 29, 1720–1741. doi: 10.1002/mds.26052
- Jones, R. S., Minogue, A. M., Connor, T. J., and Lynch, M. A. (2013). Amyloidbeta-induced astrocytic phagocytosis is mediated by CD36, CD47 and RAGE. J. Neuroimmune. Pharmacol. 8, 301–311. doi: 10.1007/s11481-012-9427-3
- Jurewicz, A., Matysiak, M., Tybor, K., Kilianek, L., Raine, C. S., and Selmaj, K. (2005). Tumour necrosis factor-induced death of adult human oligodendrocytes is mediated by apoptosis inducing factor. *Brain* 128, 2675– 2688. doi: 10.1093/brain/awh627
- Kahle, P. J., Neumann, M., Ozmen, L., Müller, V., Jacobsen, H., Schindzielorz, A., et al. (2000). Subcellular localization of wild-type and Parkinson's disease-associated mutant α-synuclein in human and transgenic mouse brain. *J. Neurosci.* 20, 6365–6373. doi: 10.1523/jneurosci.20-17-06365.2000
- Kahle, P. J., Neumann, M., Ozmen, L., Muller, V., Jacobsen, H., Spooren, W., et al. (2002). Hyperphosphorylation and insolubility of alpha-synuclein in transgenic mouse oligodendrocytes. *EMBO Rep.* 3, 583–588. doi: 10.1093/embo-reports/ kvf109
- Kalia, L. V., and Lang, A. E. (2015). Parkinson's disease. *Lancet* 386, 896–912. doi: 10.1016/s0140-6736(14)61393-3
- Kim, C., Ho, D. H., Suk, J. E., You, S., Michael, S., Kang, J., et al. (2013). Neuron-released oligomeric alpha-synuclein is an endogenous agonist of TLR2 for paracrine activation of microglia. *Nat. Commun.* 4:1562. doi: 10.1038/ ncomms2534
- Kim, C., Lee, H. J., Masliah, E., and Lee, S. J. (2016). Non-cell-autonomous neurotoxicity of alpha-synuclein through microglial toll-like receptor 2. *Exp. Neurobiol.* 25, 113–119. doi: 10.5607/en.2016.25.3.113
- Kim, T. D., Paik, S. R., and Yang, C. H. (2002). Structural and functional implications of C-terminal regions of alpha-synuclein. *Biochemistry* 41, 13782– 13790. doi: 10.1021/bi026284c
- Kim, W. S., Kagedal, K., and Halliday, G. M. (2014). Alpha-synuclein biology in Lewy body diseases. *Alzheimers Res. Ther.* 6:73. doi: 10.1186/s13195-014-0 073-2

- Kisos, H., Pukass, K., Ben-Hur, T., Richter-Landsberg, C., and Sharon, R. (2012). Increased neuronal alpha-synuclein pathology associates with its accumulation in oligodendrocytes in mice modeling alpha-synucleinopathies. *PLoS One* 7:e46817. doi: 10.1371/journal.pone.0046817
- Klegeris, A., Pelech, S., Giasson, B. I., Maguire, J., Zhang, H., McGeer, E. G., et al. (2008). Alpha-synuclein activates stress signaling protein kinases in THP-1 cells and microglia. *Neurobiol. Aging* 29, 739–752. doi: 10.1016/j.neurobiolaging. 2006.11.013
- Kohman, R. A., Bhattacharya, T. K., Kilby, C., Bucko, P., and Rhodes, J. S. (2013). Effects of minocycline on spatial learning, hippocampal neurogenesis and microglia in aged and adult mice. *Behav. Brain Res.* 242, 17–24. doi: 10. 1016/j.bbr.2012.12.032
- Komine, O., Yamashita, H., Fujimori-Tonou, N., Koike, M., Jin, S., Moriwaki, Y., et al. (2018). Innate immune adaptor TRIF deficiency accelerates disease progression of ALS mice with accumulation of aberrantly activated astrocytes. *Cell Death Differ*. 25, 2130–2146. doi: 10.1038/s41418-018-0098-3
- Konno, M., Hasegawa, T., Baba, T., Miura, E., Sugeno, N., Kikuchi, A., et al. (2012). Suppression of dynamin GTPase decreases alpha-synuclein uptake by neuronal and oligodendroglial cells: a potent therapeutic target for synucleinopathy. *Mol. Neurodegener*. 7:38. doi: 10.1186/1750-1326-7-38
- Kordower, J. H., Chu, Y., Hauser, R. A., Freeman, T. B., and Olanow, C. W. (2008). Lewy body-like pathology in long-term embryonic nigral transplants in Parkinson's disease. *Nat. Med.* 14, 504–506. doi: 10.1038/nm 1747
- Kreutzberg, G. W. (1996). Microglia: a sensor for pathological events in the CNS. *Trends Neurosci.* 19, 312–318. doi: 10.1016/0166-2236(96)10049-7
- Kuhn, S., Gritti, L., Crooks, D., and Dombrowski, Y. (2019). Oligodendrocytes in development, myelin generation and beyond. *Cells* 8:1424. doi: 10.3390/ cells8111424
- Lastres-Becker, I., Ulusoy, A., Innamorato, N. G., Sahin, G., Rabano, A., Kirik, D., et al. (2012). alpha-Synuclein expression and Nrf2 deficiency cooperate to aggravate protein aggregation, neuronal death and inflammation in earlystage Parkinson's disease. *Hum. Mol. Genet.* 21, 3173–3192. doi: 10.1093/hmg/ dds143
- Lawrence, T. (2009). The nuclear factor NF-kappaB pathway in inflammation. Cold Spring Harb. Perspect. Biol. 1:a001651. doi: 10.1101/cshperspect.a0 01651
- Leandrou, E., Emmanouilidou, E., and Vekrellis, K. (2019). Voltage-gated calcium channels and alpha-synuclein: implications in Parkinson's disease. *Front. Mol. Neurosci.* 12:237. doi: 10.3389/fnmol.2019.00237
- Lee, H. J., Kim, C., and Lee, S. J. (2010a). Alpha-synuclein stimulation of astrocytes: potential role for neuroinflammation and neuroprotection. Oxid. Med. Cell. Longev. 3, 283–287. doi: 10.4161/oxim.3.4.12809
- Lee, H. J., Suk, J. E., Patrick, C., Bae, E. J., Cho, J. H., Rho, S., et al. (2010b). Direct transfer of alpha-synuclein from neuron to astroglia causes inflammatory responses in synucleinopathies. J. Biol. Chem. 285, 9262–9272. doi: 10.1074/jbc. M109.081125
- Lee, S. B., Park, S. M., Ahn, K. J., Chung, K. C., Paik, S. R., and Kim, J. (2009). Identification of the amino acid sequence motif of alpha-synuclein responsible for macrophage activation. *Biochem. Biophys. Res. Commun.* 381, 39–43. doi: 10.1016/j.bbrc.2009.02.002
- Lee, S. J., Desplats, P., Lee, H. J., Spencer, B., and Masliah, E. (2012). Cell-to-cell transmission of alpha-synuclein aggregates. *Methods Mol. Biol.* 849, 347–359. doi: 10.1007/978-1-61779-551-0_23
- Lee, S.-J., Jeon, H., and Kandror, K. V. (2008). Alpha-synuclein is localized in a subpopulation of rat brain synaptic vesicles. Acta Neurobiol. Exp. 68, 509–515.
- Lehnardt, S., Lachance, C., Patrizi, S., Lefebvre, S., Follett, P. L., Jensen, F. E., et al. (2002). The toll-like receptor TLR4 is necessary for lipopolysaccharideinduced oligodendrocyte injury in the CNS. J. Neurosci. 22, 2478–2486. doi: 10.1523/jneurosci.22-07-02478.2002
- Levine, J. M., Reynolds, R., and Fawcett, J. W. (2001). The oligodendrocyte precursor cell in health and disease. *Trends Neurosci.* 24, 39–47. doi: 10.1016/ s0166-2236(00)01691-x
- Li, J. Y., Englund, E., Holton, J. L., Soulet, D., Hagell, P., Lees, A. J., et al. (2008). Lewy bodies in grafted neurons in subjects with Parkinson's disease suggest host-to-graft disease propagation. *Nat. Med.* 14, 501–503. doi: 10.1038/nm1746
- Li, Q., and Haney, M. S. (2020). The role of glia in protein aggregation. Neurobiol. Dis. 143:105015. doi: 10.1016/j.nbd.2020.105015

- Li, Y., Du, X. F., Liu, C. S., Wen, Z. L., and Du, J. L. (2012). Reciprocal regulation between resting microglial dynamics and neuronal activity in vivo. *Dev. Cell.* 23, 1189–1202. doi: 10.1016/j.devcel.2012.10.027
- Liddelow, S. A., and Barres, B. A. (2017). Reactive Astrocytes: production, function, and therapeutic potential. *Immunity* 46, 957–967. doi: 10.1016/j.immuni.2017. 06.006
- Liddelow, S. A., Guttenplan, K. A., Clarke, L. E., Bennett, F. C., Bohlen, C. J., Schirmer, L., et al. (2017). Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 541, 481–487. doi: 10.1038/nature21029
- Lin, K. J., Lin, K. L., Chen, S. D., Liou, C. W., Chuang, Y. C., Lin, H. Y., et al. (2019). The overcrowded crossroads: mitochondria, alpha-synuclein, and the endolysosomal system interaction in Parkinson's disease. *Int. J. Mol. Sci.* 20:5312. doi: 10.3390/ijms20215312
- Lin, W., Lin, Y., Li, J., Fenstermaker, A. G., Way, S. W., Clayton, B., et al. (2013). Oligodendrocyte-specific activation of PERK signaling protects mice against experimental autoimmune encephalomyelitis. *J. Neurosci.* 33, 5980–5991. doi: 10.1523/JNEUROSCI.1636-12.2013
- Lin, Y., Jamison, S., and Lin, W. (2012). Interferon-gamma activates nuclear factorkappa B in oligodendrocytes through a process mediated by the unfolded protein response. *PLoS One* 7:e36408. doi: 10.1371/journal.pone.0036408
- Lindstrom, V., Gustafsson, G., Sanders, L. H., Howlett, E. H., Sigvardson, J., Kasrayan, A., et al. (2017). Extensive uptake of alpha-synuclein oligomers in astrocytes results in sustained intracellular deposits and mitochondrial damage. *Mol. Cell. Neurosci.* 82, 143–156. doi: 10.1016/j.mcn.2017.04.009
- Liu, G., Zhang, C., Yin, J., Li, X., Cheng, F., Li, Y., et al. (2009). alpha-Synuclein is differentially expressed in mitochondria from different rat brain regions and dose-dependently down-regulates complex I activity. *Neurosci. Lett.* 454, 187–192. doi: 10.1016/j.neulet.2009.02.056
- Liu, M., Qin, L., Wang, L., Tan, J., Zhang, H., Tang, J., et al. (2018). alphasynuclein induces apoptosis of astrocytes by causing dysfunction of the endoplasmic reticulumGolgi compartment. *Mol. Med. Rep.* 18, 322–332. doi: 10.3892/mmr. 2018.9002
- Liu, W., Tang, Y., and Feng, J. (2011). Cross talk between activation of microglia and astrocytes in pathological conditions in the central nervous system. *Life Sci.* 89, 141–146. doi: 10.1016/j.lfs.2011.05.011
- Lopez-Armada, M. J., Riveiro-Naveira, R. R., Vaamonde-Garcia, C., and Valcarcel-Ares, M. N. (2013). Mitochondrial dysfunction and the inflammatory response. *Mitochondrion* 13, 106–118. doi: 10.1016/j.mito.2013.01.003
- Lopez-Cuina, M., Foubert-Samier, A., Tison, F., and Meissner, W. G. (2018). Present and future of disease-modifying therapies in multiple system atrophy. *Auton. Neurosci.* 211, 31–38. doi: 10.1016/j.autneu.2017.12.008
- Luth, E. S., Stavrovskaya, I. G., Bartels, T., Kristal, B. S., and Selkoe, D. J. (2014). Soluble, prefibrillar alpha-synuclein oligomers promote complex I-dependent, Ca2+-induced mitochondrial dysfunction. J. Biol. Chem. 289, 21490–21507. doi: 10.1074/jbc.M113.545749
- Maiza, A., Chantepie, S., Vera, C., Fifre, A., Huynh, M. B., Stettler, O., et al. (2018). The role of heparan sulfates in protein aggregation and their potential impact on neurodegeneration. *FEBS Lett.* 592, 3806–3818. doi: 10.1002/1873-3468.13082
- Marrìn-Teva, J. L., Dusart, I., Colin, C., Gervais, A., van Rooijen, N., and Mallat, M. (2004). Microglia promote the death of developing purkinje cells. *Neuron* 41, 535–547. doi: 10.1016/s0896-6273(04)00069-8
- Maroteaux, L., Campanelli, J. T., and Scheller, R. H. (1988). Synuclein: a neuronspecific protein localized to the nucleus and presynaptic nerve terminal. *J. Neurosci.* 8, 2804–2815. doi: 10.1523/jneurosci.08-08-02804.1988
- May, V. E., Ettle, B., Poehler, A. M., Nuber, S., Ubhi, K., Rockenstein, E., et al. (2014). alpha-Synuclein impairs oligodendrocyte progenitor maturation in multiple system atrophy. *Neurobiol. Aging* 35, 2357–2368. doi: 10.1016/j. neurobiolaging.2014.02.028
- McAvoy, K., and Kawamata, H. (2019). Glial mitochondrial function and dysfunction in health and neurodegeneration. *Mol. Cell. Neurosci.* 101:103417. doi: 10.1016/j.mcn.2019.103417
- McCann, H., Stevens, C. H., Cartwright, H., and Halliday, G. M. (2014). α-Synucleinopathy phenotypes. *Parkinsonism Relat. Disord.* 20, S62–S67. doi: 10.1016/s1353-8020(13)70017-8
- McKeith, I. G., Boeve, B. F., Dickson, D. W., Halliday, G., Taylor, J. P., Weintraub, D., et al. (2017). Diagnosis and management of dementia with Lewy bodies: fourth consensus report of the DLB consortium. *Neurology* 89, 88–100. doi: 10.1212/WNL.000000000004058

- Medeiros, G. (2000). Distribution of sulfated glycosaminoglycans in the animal kingdom: widespread occurrence of heparin-like compounds in invertebrates. *Biochim. Biophys. Acta Gen. Subj.* 1475, 287–294. doi: 10.1016/s0304-4165(00) 00079-9
- Melachroinou, K., Xilouri, M., Emmanouilidou, E., Masgrau, R., Papazafiri, P., Stefanis, L., et al. (2013). Deregulation of calcium homeostasis mediates secreted alpha-synuclein-induced neurotoxicity. *Neurobiol. Aging* 34, 2853–2865. doi: 10.1016/j.neurobiolaging.2013.06.006
- Melo, T. Q., Copray, S. J., and Ferrari, M. F. R. (2018). Alpha-synuclein toxicity on protein quality control, mitochondria and endoplasmic reticulum. *Neurochem. Res.* 43, 2212–2223. doi: 10.1007/s11064-018-2673-x
- Meredith, G. E., Totterdell, S., Petroske, E., Santa Cruz, K., Callison, R. C., and Lau, Y. S. (2002). Lysosomal malfunction accompanies alpha-synuclein aggregation in a progressive mouse model of Parkinson's disease. *Brain Res.* 956, 156–165. doi: 10.1016/s0006-8993(02)03514-x
- Miller, D. W., Johnson, J. M., Solano, S. M., Hollingsworth, Z. R., Standaert, D. G., Young, A. B., et al. (2005). Absence of alpha-synuclein mRNA expression in normal and multiple system atrophy oligodendroglia. *J. Neural Transm.* 112, 1613–1624. doi: 10.1007/s00702-005-0378-1
- Mirza, B., Hadberg, H., Thomsen, P., and Moos, T. (1999). The absence of reactive astrocytosis is indicative of a unique inflammatory process in Parkinson's disease. *Neuroscience* 95, 425–432. doi: 10.1016/s0306-4522(99)00455-8
- Moehle, M. S., and West, A. B. (2015). M1 and M2 immune activation in Parkinson's disease: foe and ally? *Neuroscience* 302, 59–73. doi: 10.1016/j. neuroscience.2014.11.018
- Mogi, M., Harada, M., Kondo, T., Riederer, P., Inagaki, H., Minami, M., et al. (1994a). Interleukin-1 β , interleukin-6, epidermal growth factor and transforming growth factor- α are elevated in the brain from parkinsonian patients. *Neurosci. Lett.* 180, 147–150. doi: 10.1016/0304-3940(94)90508-8
- Mogi, M., Harada, M., Riederer, P., Narabayashi, H., Fujita, K., and Nagatsu, T. (1994b). Tumor necrosis factor-α (TNF-α) increases both in the brain and in the cerebrospinal fluid from parkinsonian patients. *Neurosci. Lett.* 165, 208–210. doi: 10.1016/0304-3940(94)90746-3
- Monzio Compagnoni, G., and Di Fonzo, A. (2019). Understanding the pathogenesis of multiple system atrophy: state of the art and future perspectives. *Acta Neuropathol. Commun.* 7:113. doi: 10.1186/s40478-019-0730-6
- Mori, A., Imai, Y., and Hattori, N. (2020). Lipids: key players that modulate alphasynuclein toxicity and neurodegeneration in Parkinson's disease. *Int. J. Mol. Sci.* 21:3301. doi: 10.3390/ijms21093301
- Mori, F., Tanji, K., Yoshimoto, M., Takahashi, H., and Wakabayashi, K. (2002). Demonstration of alpha-synuclein immunoreactivity in neuronal and glial cytoplasm in normal human brain tissue using proteinase K and formic acid pretreatment. *Exp. Neurol.* 176, 98–104. doi: 10.1006/exnr.2002.7929
- Morizawa, Y. M., Hirayama, Y., Ohno, N., Shibata, S., Shigetomi, E., Sui, Y., et al. (2017). Reactive astrocytes function as phagocytes after brain ischemia via ABCA1-mediated pathway. *Nat. Commun.* 8:28. doi: 10.1038/s41467-017-00037-1
- Motamedi-Shad, N., Monsellier, E., Torrassa, S., Relini, A., and Chiti, F. (2009). Kinetic analysis of amyloid formation in the presence of heparan sulfate: faster unfolding and change of pathway. *J. Biol. Chem.* 284, 29921–29934. doi: 10. 1074/jbc.M109.018747
- Muller, T., Blum-Degen, D., Przuntek, H., and Kuhn, W. (1998). Interleukin-6 levels in cerebrospinal fluid inversely correlate to severity of Parkinson's disease. Acta Neurol. Scand. 98, 142–144. doi: 10.1111/j.1600-0404.1998.tb01 736.x
- Multiple-System Atrophy Research Collaboration (2013). Mutations in COQ2 in familial and sporadic multiple-system atrophy. *N. Engl. J. Med.* 369, 233–244. doi: 10.1056/NEJMoa1212115
- Muppidi, S., and Miglis, M. G. (2017). Is pure autonomic failure an early marker for Parkinson disease, dementia with Lewy bodies, and multiple system atrophy? And other updates on recent autonomic research. *Clin. Auton. Res.* 27, 71–73. doi: 10.1007/s10286-017-0408-8
- Mythri, R. B., Venkateshappa, C., Harish, G., Mahadevan, A., Muthane, U. B., Yasha, T. C., et al. (2011). Evaluation of markers of oxidative stress, antioxidant function and astrocytic proliferation in the striatum and frontal cortex of Parkinson's disease brains. *Neurochem. Res.* 36, 1452–1463. doi: 10.1007/ s11064-011-0471-9

- Nath, S., Goodwin, J., Engelborghs, Y., and Pountney, D. L. (2011). Raised calcium promotes alpha-synuclein aggregate formation. *Mol Cell. Neurosci.* 46, 516– 526. doi: 10.1016/j.mcn.2010.12.004
- Neumann, H., Kotter, M. R., and Franklin, R. J. (2009). Debris clearance by microglia: an essential link between degeneration and regeneration. *Brain* 132, 288–295. doi: 10.1093/brain/awn109
- Nickols, J. C., Valentine, W., Kanwal, S., and Carter, B. D. (2003). Activation of the transcription factor NF-kappaB in Schwann cells is required for peripheral myelin formation. *Nat. Neurosci.* 6, 161–167. doi: 10.1038/nn995
- No authors listed (1996). Consensus statement on the definition of orthostatic hypotension, pure autonomic failure, and multiple system atrophy. The Consensus Committee of the American Autonomic Society and the American Academy of Neurology. *Neurology* 46:1470. doi: 10.1212/wnl.46.5.1470
- Olson, J. K., and Miller, S. D. (2004). Microglia initiate central nervous system innate and adaptive immune responses through multiple TLRs. *J. Immunol.* 173, 3916–3924. doi: 10.4049/jimmunol.173.6.3916
- Orme, T., Guerreiro, R., and Bras, J. (2018). The genetics of dementia with lewy bodies: current understanding and future directions. *Curr. Neurol. Neurosci. Rep.* 18:67. doi: 10.1007/s11910-018-0874-y
- Pajares, M., Rojo, A. I., Manda, G., Bosca, L., and Cuadrado, A. (2020). Inflammation in Parkinson's disease: mechanisms and therapeutic implications. *Cells* 9:1687. doi: 10.3390/cells9071687
- Pan, W., Stone, K. P., Hsuchou, H., Manda, V. K., Zhang, Y., and Kastin, A. J. (2011). Cytokine signaling modulates blood-brain barrier function. *Curr. Pharm. Des.* 17, 3729–3740. doi: 10.2174/138161211798220918
- Park, J. S., Davis, R. L., and Sue, C. M. (2018). Mitochondrial dysfunction in Parkinson's disease: new mechanistic insights and therapeutic perspectives. *Curr. Neurol. Neurosci. Rep.* 18:21. doi: 10.1007/s11910-018-0829-3
- Parker, W. D. Jr., Parks, J. K., and Swerdlow, R. H. (2008). Complex I deficiency in Parkinson's disease frontal cortex. *Brain Res.* 1189, 215–218. doi: 10.1016/j. brainres.2007.10.061
- Peferoen, L., Kipp, M., van der Valk, P., van Noort, J. M., and Amor, S. (2014). Oligodendrocyte-microglia cross-talk in the central nervous system. *Immunology* 141, 302–313. doi: 10.1111/imm.12163
- Perry, E. K., Irving, D., Kerwin, J. M., McKeith, I. G., Thompson, P., Collerton, D., et al. (1993). Cholinergic transmitter and neurotrophic activities in Lewy body dementia: similarity to Parkinson's and distinction from Alzheimer disease. *Alzheimer Dis. Assoc. Disord.* 7, 69–79. doi: 10.1097/00002093-199307020-00002
- Piao, Y. S., Hayashi, S., Hasegawa, M., Wakabayashi, K., Yamada, M., Yoshimoto, M., et al. (2001). Co-localization of alpha-synuclein and phosphorylated tau in neuronal and glial cytoplasmic inclusions in a patient with multiple system atrophy of long duration. *Acta Neuropathol.* 101, 285–293. doi: 10.1007/ s004010000292
- Pozo Devoto, V. M., and Falzone, T. L. (2017). Mitochondrial dynamics in Parkinson's disease: a role for alpha-synuclein? *Dis. Model. Mech.* 10, 1075– 1087.
- Pukass, K., Goldbaum, O., and Richter-Landsberg, C. (2015). Mitochondrial impairment and oxidative stress compromise autophagosomal degradation of alpha-synuclein in oligodendroglial cells. J. Neurochem. 135, 194–205. doi: 10. 1111/jnc.13256
- Quinzii, C. M., and Hirano, M. (2010). Coenzyme Q and mitochondrial disease. Dev. Disabil. Res. Rev. 16, 183–188. doi: 10.1002/ddrr.108
- Ramirez, G., Toro, R., Dobeli, H., and von Bernhardi, R. (2005). Protection of rat primary hippocampal cultures from A beta cytotoxicity by pro-inflammatory molecules is mediated by astrocytes. *Neurobiol. Dis.* 19, 243–254. doi: 10.1016/ j.nbd.2005.01.007
- Rannikko, E. H., Weber, S. S., and Kahle, P. J. (2015). Exogenous alpha-synuclein induces toll-like receptor 4 dependent inflammatory responses in astrocytes. *BMC Neurosci.* 16:57. doi: 10.1186/s12868-015-0192-0
- Rapp, M. A., Schnaider-Beeri, M., Wysocki, M., Guerrero-Berroa, E., Grossman, H. T., Heinz, A., et al. (2011). Cognitive decline in patients with dementia as a function of depression. *Am. J. Geriatr. Psychiatry* 19, 357–363. doi: 10.1097/ JGP.0b013e3181e898d0
- Reale, M., Greig, N. H., and Kamal, M. A. (2009). Peripheral chemo-cytokine profiles in Alzheimers and Parkinson's diseases. *Mini. Rev. Med. Chem.* 9, 1229–1241. doi: 10.2174/138955709789055199

- Reyes, J. F., Olsson, T. T., Lamberts, J. T., Devine, M. J., Kunath, T., and Brundin, P. (2015). A cell culture model for monitoring alpha-synuclein cell-to-cell transfer. *Neurobiol. Dis.* 77, 266–275. doi: 10.1016/j.nbd.2014.07.003
- Rocha, E. M., De Miranda, B., and Sanders, L. H. (2018). Alpha-synuclein: pathology, mitochondrial dysfunction and neuroinflammation in Parkinson's disease. *Neurobiol. Dis.* 109, 249–257. doi: 10.1016/j.nbd.2017.04.004
- Rojanathammanee, L., Murphy, E. J., and Combs, C. K. (2011). Expression of mutant alpha-synuclein modulates microglial phenotype in vitro. *J. Neuroinflammation* 8:44. doi: 10.1186/1742-2094-8-44
- Roodveldt, C., Labrador-Garrido, A., Gonzalez-Rey, E., Lachaud, C. C., Guilliams, T., Fernandez-Montesinos, R., et al. (2013). Preconditioning of microglia by alpha-synuclein strongly affects the response induced by toll-like receptor (TLR) stimulation. *PLoS One* 8:e79160. doi: 10.1371/journal.pone.0079160
- Rostami, J., Holmqvist, S., Lindstrom, V., Sigvardson, J., Westermark, G. T., Ingelsson, M., et al. (2017). Human astrocytes transfer aggregated alphasynuclein via tunneling nanotubes. *J. Neurosci.* 37, 11835–11853. doi: 10.1523/ JNEUROSCI.0983-17.2017
- Rusnakova, V., Honsa, P., Dzamba, D., Stahlberg, A., Kubista, M., and Anderova, M. (2013). Heterogeneity of astrocytes: from development to injury - single cell gene expression. *PLoS One* 8:e69734. doi: 10.1371/journal.pone.0069734
- Sala, G., Marinig, D., Arosio, A., and Ferrarese, C. (2016). Role of chaperonemediated autophagy dysfunctions in the pathogenesis of Parkinson's disease. *Front. Mol. Neurosci.* 9:157. doi: 10.3389/fnmol.2016.00157
- Schapira, A. H. V., Cooper, J. M., Dexter, D., Jenner, P., Clark, J. B., and Marsden, C. D. (1989). Mitochondrial complex I deficiency in Parkinson's disease. *Lancet* 333:1269. doi: 10.1016/s0140-6736(89)92366-0
- Schwarz, L., Goldbaum, O., Bergmann, M., Probst-Cousin, S., and Richter-Landsberg, C. (2012). Involvement of macroautophagy in multiple system atrophy and protein aggregate formation in oligodendrocytes. J. Mol. Neurosci. 47, 256–266. doi: 10.1007/s12031-012-9733-5
- Shao, Q. H., Yan, W. F., Zhang, Z., Ma, K. L., Peng, S. Y., Cao, Y. L., et al. (2019). Nurr1: a vital participant in the TLR4-NF-kappaB signal pathway stimulated by alpha-synuclein in BV-2cells. *Neuropharmacology* 144, 388–399. doi: 10.1016/j. neuropharm.2018.04.008
- Simard, M., and Nedergaard, M. (2004). The neurobiology of glia in the context of water and ion homeostasis. *Neuroscience* 129, 877–896. doi: 10.1016/j. neuroscience.2004.09.053
- Singer, W., Berini, S. E., Sandroni, P., Fealey, R. D., Coon, E. A., Suarez, M. D., et al. (2017). Pure autonomic failure: predictors of conversion to clinical CNS involvement. *Neurology* 88, 1129–1136. doi: 10.1212/WNL.00000000003737
- Sofroniew, M. V., and Vinters, H. V. (2010). Astrocytes: biology and pathology. Acta Neuropathol. 119, 7–35. doi: 10.1007/s00401-009-0619-8
- Song, Y. J., Halliday, G. M., Holton, J. L., Lashley, T., OSullivan, S. S., McCann, H., et al. (2009). Degeneration in different parkinsonian syndromes relates to astrocyte type and astrocyte protein expression. *J. Neuropathol. Exp. Neurol.* 68, 1073–1083. doi: 10.1097/NEN.0b013e3181b66f1b
- Soulet, D., and Rivest, S. (2008). Microglia. *Curr. Biol.* 18, R506–R508. doi: 10.1016/ j.cub.2008.04.047
- Spillantini, M. G., Schmidt, M. L., Lee, V. M., Trojanowski, J. Q., Jakes, R., and Goedert, M. (1997). Alpha-synuclein in Lewy bodies. *Nature* 388, 839–840. doi: 10.1038/42166
- Stefanova, N., Fellner, L., Reindl, M., Masliah, E., Poewe, W., and Wenning, G. K. (2011). Toll-like receptor 4 promotes alpha-synuclein clearance and survival of nigral dopaminergic neurons. *Am. J. Pathol.* 179, 954–963. doi: 10.1016/j.ajpath. 2011.04.013
- Stefanova, N., Klimaschewski, L., Poewe, W., Wenning, G. K., and Reindl, M. (2001). Glial cell death induced by overexpression of alpha-synuclein. *J. Neurosci. Res.* 65, 432–438. doi: 10.1002/jnr.1171
- Stefanova, N., Reindl, M., Neumann, M., Haass, C., Poewe, W., Kahle, P. J., et al. (2005). Oxidative stress in transgenic mice with oligodendroglial α-synuclein overexpression replicates the characteristic neuropathology of multiple system atrophy. Am. J. Pathol. 166, 869–876. doi: 10.1016/s0002-9440(10)62307-3
- Steiner, J. A., Quansah, E., and Brundin, P. (2018). The concept of alpha-synuclein as a prion-like protein: ten years after. *Cell Tissue Res.* 373, 161–173. doi: 10. 1007/s00441-018-2814-1
- Stone, S., Jamison, S., Yue, Y., Durose, W., Schmidt-Ullrich, R., and Lin, W. (2017). NF-kappaB activation protects oligodendrocytes against inflammation. *J. Neurosci.* 37, 9332–9344. doi: 10.1523/JNEUROSCI.1608-17.2017

- Stubendorff, K., Aarsland, D., Minthon, L., and Londos, E. (2012). The impact of autonomic dysfunction on survival in patients with dementia with Lewy bodies and Parkinson's disease with dementia. *PLoS One* 7:e45451. doi: 10.1371/ journal.pone.0045451
- Su, X., Maguire-Zeiss, K. A., Giuliano, R., Prifti, L., Venkatesh, K., and Federoff, H. J. (2008). Synuclein activates microglia in a model of Parkinson's disease. *Neurobiol. Aging* 29, 1690–1701. doi: 10.1016/j.neurobiolaging.2007. 04.006
- Subramaniam, S. R., and Federoff, H. J. (2017). Targeting microglial activation states as a therapeutic avenue in Parkinson's disease. *Front. Aging Neurosci.* 9:176. doi: 10.3389/fnagi.2017.00176
- Surendranathan, A., Kane, J. P. M., Bentley, A., Barker, S. A. H., Taylor, J. P., Thomas, A. J., et al. (2020). Clinical diagnosis of Lewy body dementia. B. J. Psychol. Open 6:e61. doi: 10.1192/bjo.2020.44
- Tang, S. C., Arumugam, T. V., Xu, X., Cheng, A., Mughal, M. R., Jo, D. G., et al. (2007). Pivotal role for neuronal Toll-like receptors in ischemic brain injury and functional deficits. *Proc. Natl. Acad. Sci. U.S.A.* 104, 13798–13803. doi: 10.1073/pnas.0702553104
- Tang, Y., and Le, W. (2016). Differential roles of M1 and M2 microglia in neurodegenerative diseases. *Mol. Neurobiol.* 53, 1181–1194. doi: 10.1007/ s12035-014-9070-5
- Tanji, K., Odagiri, S., Maruyama, A., Mori, F., Kakita, A. A., Takahashi, H., et al. (2013). Alteration of autophagosomal proteins in the brain of multiple system atrophy. *Neurobiol. Dis.* 49, 190–198. doi: 10.1016/j.nbd.2012.08.017
- Thaisetthawatkul, P. (2016). Pure autonomic failure. Curr. Neurol. Neurosci. Rep. 16:74. doi: 10.1007/s11910-016-0673-2
- Thorburne, S. K., and Juurlink, B. H. (1996). Low glutathione and high iron govern the susceptibility of oligodendroglial precursors to oxidative stress. *J. Neurochem.* 67, 1014–1022. doi: 10.1046/j.1471-4159.1996.67031014.x
- Tichauer, J., Saud, K., and von Bernhardi, R. (2007). Modulation by astrocytes of microglial cell-mediated neuroinflammation: effect on the activation of microglial signaling pathways. *Neuroimmunomodulation* 14, 168–174. doi: 10. 1159/000110642
- Tiraboschi, P., Hansen, L. A., Alford, M., Sabbagh, M. N., Schoos, B., Masliah, E., et al. (2000). Cholinergic dysfunction in diseases with Lewy bodies. *Neurology* 54, 407–411. doi: 10.1212/wnl.54.2.407
- Tjalkens, R. B., Popichak, K. A., and Kirkley, K. A. (2017). Inflammatory activation of microglia and astrocytes in manganese neurotoxicity. *Adv. Neurobiol.* 18, 159–181. doi: 10.1007/978-3-319-60189-2_8
- Tong, X., Ao, Y., Faas, G. C., Nwaobi, S. E., Xu, J., Haustein, M. D., et al. (2014). Astrocyte Kir4.1 ion channel deficits contribute to neuronal dysfunction in Huntingtons disease model mice. *Nat. Neurosci.* 17, 694–703. doi: 10.1038/nn. 3691
- Tosatto, L., Andrighetti, A. O., Plotegher, N., Antonini, V., Tessari, I., Ricci, L., et al. (2012). Alpha-synuclein pore forming activity upon membrane association. *Biochim. Biophys. Acta* 1818, 2876–2883. doi: 10.1016/j.bbamem.2012.07.007
- Tousi, B. (2017). Diagnosis and management of cognitive and behavioral changes in dementia with lewy bodies. *Curr. Treat. Options Neurol.* 19:42. doi: 10.1007/ s11940-017-0478-x
- Trancikova, A., Tsika, E., and Moore, D. J. (2012). Mitochondrial dysfunction in genetic animal models of Parkinson's disease. *Antioxid. Redox Signal.* 16, 896–919. doi: 10.1089/ars.2011.4200
- Tremblay, M. E., Cookson, M. R., and Civiero, L. (2019). Glial phagocytic clearance in Parkinson's disease. *Mol. Neurodegener.* 14:16. doi: 10.1186/s13024-019-0314-8
- Ubhi, K., Low, P., and Masliah, E. (2011). Multiple system atrophy: a clinical and neuropathological perspective. *Trends Neurosci.* 34, 581–590. doi: 10.1016/j. tins.2011.08.003
- Ueda, K., Fukushima, H., Masliah, E., Xia, Y., Iwai, A., Yoshimoto, M., et al. (1993). Molecular cloning of cDNA encoding an unrecognized component of amyloid in Alzheimer disease. *Proc. Natl. Acad. Sci. U.S.A.* 90, 11282–11286. doi: 10.1073/pnas.90.23.11282
- Velayudhan, L., Ffytche, D., Ballard, C., and Aarsland, D. (2017). New therapeutic strategies for lewy body dementias. *Curr. Neurol. Neurosci. Rep.* 17:68. doi: 10.1007/s11910-017-0778-2
- Venezia, S., Refolo, V., Polissidis, A., Stefanis, L., Wenning, G. K., and Stefanova, N. (2017). Toll-like receptor 4 stimulation with monophosphoryl lipid A ameliorates motor deficits and nigral neurodegeneration triggered by

extraneuronal alpha-synucleinopathy. Mol. Neurodegener. 12:52. doi: 10.1186/ s13024-017-0195-7

- Vicario, M., Cieri, D., Brini, M., and Cali, T. (2018). The close encounter between alpha-synuclein and mitochondria. *Front. Neurosci.* 12:388. doi: 10.3389/fnins. 2018.00388
- Vila, M., Jackson-Lewis, V., Guegan, C., Wu, D. C., Teismann, P., Choi, D. K., et al. (2001). The role of glial cells in Parkinson's disease. *Curr. Opin. Neurol.* 14, 483–489. doi: 10.1097/00019052-200108000-00009
- von Bartheld, C. S., Bahney, J., and Herculano-Houzel, S. (2016). The search for true numbers of neurons and glial cells in the human brain: a review of 150 years of cell counting. *J. Comp. Neurol.* 524, 3865–3895. doi: 10.1002/cne.24040
- Wakabayashi, K., Hayashi, S., Yoshimoto, M., Kudo, H., and Takahashi, H. (2000). NACP/alpha-synuclein-positive filamentous inclusions in astrocytes and oligodendrocytes of Parkinson's disease brains. *Acta Neuropathol.* 99, 14–20. doi: 10.1007/pl00007400
- Walker, Z., Possin, K. L., Boeve, B. F., and Aarsland, D. (2015). Lewy body dementias. *Lancet* 386, 1683–1697. doi: 10.1016/s0140-6736(15)00462-6
- Walter, L., and Neumann, H. (2009). Role of microglia in neuronal degeneration and regeneration. *Semin. Immunopathol.* 31, 513–525. doi: 10.1007/s00281-009-0180-5
- Wang, L., Das, U., Scott, D. A., Tang, Y., McLean, P. J., and Roy, S. (2014). alphasynuclein multimers cluster synaptic vesicles and attenuate recycling. *Curr. Biol.* 24, 2319–2326. doi: 10.1016/j.cub.2014.08.027
- Wang, Q., Liu, Y., and Zhou, J. (2015). Neuroinflammation in Parkinson's disease and its potential as therapeutic target. *Transl. Neurodegener.* 4:19. doi: 10.1186/ s40035-015-0042-0
- Wang, W., Perovic, I., Chittuluru, J., Kaganovich, A., Nguyen, L. T., Liao, J., et al. (2011). A soluble alpha-synuclein construct forms a dynamic tetramer. *Proc. Natl. Acad. Sci. U.S.A.* 108, 17797–17802. doi: 10.1073/pnas.1113260108
- Wang, X. J., Ma, M. M., Zhou, L. B., Jiang, X. Y., Hao, M. M., Teng, R. K. F., et al. (2020). Autonomic ganglionic injection of alpha-synuclein fibrils as a model of pure autonomic failure alpha-synucleinopathy. *Nat. Commun.* 11:934. doi: 10.1038/s41467-019-14189-9
- Wang, X. L., Feng, S. T., Wang, Z. Z., Yuan, Y. H., Chen, N. H., and Zhang, Y. (2020). Parkin, an E3 ubiquitin ligase, plays an essential role in mitochondrial quality control in Parkinson's disease. *Cell. Mol. Neurobiol.* [Epub ahead of print]. doi: 10.1007/s10571-020-00914-912
- Wilkins, A., Majed, H., Layfield, R., Compston, A., and Chandran, S. (2003). Oligodendrocytes promote neuronal survival and axonal length by distinct intracellular mechanisms: a novel role for oligodendrocyte-derived glial cell line-derived neurotrophic factor. J. Neurosci. 23, 4967–4974. doi: 10.1523/ jneurosci.23-12-04967.2003

- Woerman, A. L., Watts, J. C., Aoyagi, A., Giles, K., Middleton, L. T., and Prusiner, S. B. (2018). alpha-synuclein: multiple system atrophy prions. *Cold Spring Harb. Perspect. Med.* 8:a024588. doi: 10.1101/cshperspect.a0 24588
- Wojda, U., Salinska, E., and Kuznicki, J. (2008). Calcium ions in neuronal degeneration. *IUBMB Life* 60, 575–590. doi: 10.1002/iub.91
- Yan, X., Wang, B., Hu, Y., Wang, S., and Zhang, X. (2020). Abnormal mitochondrial quality control in neurodegenerative diseases. *Front. Cell. Neurosci.* 14:138. doi: 10.3389/fncel.2020.00138
- Yang, D., Elner, S. G., Bian, Z. M., Till, G. O., Petty, H. R., and Elner, V. M. (2007). Pro-inflammatory cytokines increase reactive oxygen species through mitochondria and NADPH oxidase in cultured RPE cells. *Exp. Eye Res.* 85, 462–472. doi: 10.1016/j.exer.2007.06.013
- Yang, J., Zhao, Z., Gu, M., Feng, X., and Xu, H. (2019). Release and uptake mechanisms of vesicular Ca(2+) stores. *Protein Cell* 10, 8–19. doi: 10.1007/s13238-018-0523-x
- Yang, Y., Rong, X., Lv, X., Jiang, W., Yang, Y., Lai, D., et al. (2017). Inhibition of mevalonate pathway prevents ischemia-induced cardiac dysfunction in rats via RhoA-independent signaling pathway. *Cardiovasc. Ther.* 35:e12285. doi: 10.1111/1755-5922.12285
- Yoon, Y. S., Ahn, W. J., Ricarte, D., Ortiz, D., Shin, C. Y., Lee, S. J., et al. (2020). Alpha-synuclein inclusion formation in human oligodendrocytes. *Biomol. Ther.* [Epub ahead of print]. doi: 10.4062/biomolther.2020.081
- Yu, W. W., Cao, S. N., Zang, C. X., Wang, L., Yang, H. Y., Bao, X. Q., et al. (2018). Heat shock protein 70 suppresses neuroinflammation induced by alphasynuclein in astrocytes. *Mol. Cell. Neurosci.* 86, 58–64. doi: 10.1016/j.mcn.2017. 11.013
- Zhang, W., Wang, T., Pei, Z., Miller, D. S., Wu, X., Block, M. L., et al. (2005). Aggregated alpha-synuclein activates microglia: a process leading to disease progression in Parkinson's disease. *FASEB J.* 19, 533–542. doi: 10.1096/fj.04-2751com

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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